



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : A61K	A2	(11) International Publication Number: WO 98/33470
		(43) International Publication Date: 6 August 1998 (06.08.98)

(21) International Application Number: PCT/US98/00235

(22) International Filing Date: 9 January 1998 (09.01.98)

(30) Priority Data:
08/782,048 10 January 1997 (10.01.97) US

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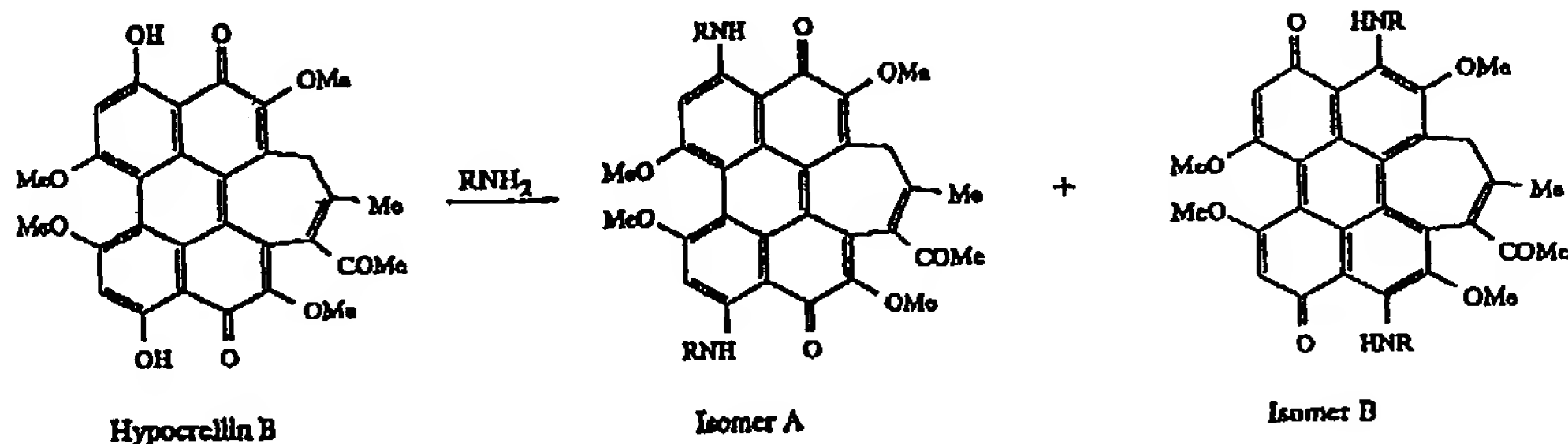
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(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

Without international search report and to be republished upon receipt of that report.

(54) Title: SUBSTITUTED PERYLENEQUINONES FOR USE IN PHOTODYNAMIC THERAPY



(57) Abstract

The invention involves a method and compositions for use in photodynamic therapy. Novel perylenequinone derivatives, conjugates comprising perylenequinone derivatives and a binding agent, and methods of treatment using these compositions are disclosed.

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SUBSTITUTED PERYLENEQUINONES FOR USE IN PHOTODYNAMIC THERAPY

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Technical Field of the Invention

The invention involves a method for using and enhancing the activity of compounds that have anti-cancer activity, anti-viral activity, cosmetic improvement, and/or non-malignant disease activity in animals, including humans. The invention
11 also involves diagnostic methods and compositions.

Background of the Invention

Treatment for cancer has traditionally encompassed three main strategies: surgery, chemotherapy, and radiotherapy. Although considerable progress in these
16 areas has been attained, the search for more effective and safe alternative treatments continues. Lipson, et al. were the first to use photodynamic therapy (PDT), in 1966 at the Mayo Clinic [*Proc. IX Internat. Cancer Congress*, page 393 (1966)]. By 1972 widespread interest in PDT had prompted animal experiments, the results of which demonstrated that the combination of a photosensitizer (e.g., fluorescein), light, and
21 cellular oxygen could inhibit tumor growth [Dougherty, T.J., *JNCI* 52:1333 (1974)]. The first animal study using hematoporphyrin derivatives (HpD) was reported in 1975 [Dougherty, et al., *JNCI* 55:115 (1975)].

Since the advent of HpD and its more purified version, Photofrin®, PDT of tumors has progressed to phase III clinical trials. Although HpD has been a useful
26 tissue photosensitizer, problems are associated with its use, including prolonged cutaneous phototoxicity; the compositions are oligomeric mixtures of hydrophilic molecules prone to molecular aggregation (with concomitant loss of photopotential); complicated pharmacokinetics; poor absorption and photoactivation in the "therapeutic window" (600 nm to 850 nm, i.e., visible red light). Furthermore, its batch

1 reproducibility, even the clinical compositions, has been poor.

Interest in PDT to treat human malignancies and non-malignant conditions has increased markedly since the advent and clinical application of hematoporphyrin derivatives. To date PDT of cancers (in both laboratory and clinical trials) has primarily involved the use of porphyrin-based photosensitizers e.g., Photofrin-II®.

6 These compounds have been used, and interest in PDT has increased, despite porphyrin-based photosensitizer's sub-optimal light absorption characteristics, source-dependent biological response and molecular composition, and difficulties associated with prolonged photosensitization of the host.

These limitations have prompted a search for alternative photosensitizers.

11 Current candidates include benzoporphyrin derivative monoacid ring A [Richter, et al., *Photochem. Photobiol.* 52:495 (1990)], and 5-aminolevulinic acid/protoporphyrin IX [Kennedy, et al., *J. Photochem. Photobiol B Biol.* 6(1-2):143 (1990)]. Additional studies include merocyanine 540, phthalocyanine (CASPc), mono-L-aspartyl chlorin e₆ (MACE), Nile blue, anthrapyrroles, anthracenediones, anthracyclines, and
16 hypocrellins [Diwu, et al., *Photochem. Photobiol.* 52(3):609 (1990)].

The photosensitizing properties of perylenequinoid pigments (PQPs), such as hypocrellins, in biological systems have been recognized during the past two decades. See Diwu, et al., *J. Photochem. Photobiol. A: Chem.*, 64:273 (1992); Zhang et al., (1989); and Wan, et al., "Hypocrellin A, a new drug for photochemotherapy," *Kexue Tongbao*
21 (English edition) 26:1040 (1981).

Hypocrellins derive their name from *Hypocrella bambusae* sacc., a parasitic fungus of the Sinarundinaria species, which grows abundantly in the northwestern region of the Yunnan Province (People's Republic of China), the southeastern region of Tibet, and certain parts of Sri Lanka. Hypocrellins belong to the general class of
26 perylenequinoid pigments, and include hypocrellin A (HA) and hypocrellin B (HB).

Oral administration of HA causes hypericisms, a state of skin sensitivity to visible light. It has now been shown that the photosensitizing effects of HA depend on the presence of oxygen, indicating that naturally occurring HA can be used as a photodynamic agent (Wan, et al., above).

Summary of the Invention

In accordance with the present invention, derivatives of perylenequinone pigments (PQPs) are a new class of photosensitizing agents, and are useful in photodynamic therapy. Moreover, PQP derivatives, particularly functionalized PQPs, may be conjugated to binding agents that bind pre-determined cells or structures *in vitro* or *in vivo*.

The methods and compositions of the present invention provide meaningful improvements over Photofrin®. For example, hypocrellins exhibit substantial absorption in the red spectral region; produce high singlet oxygen yield; can be produced in pure, monomeric form; may be derivatized to optimize properties of red light absorption, tissue biodistribution, and toxicity; have reduced residual cutaneous photosensitivity; and are rapidly excreted. They afford nuclear targeting by covalent attachment to DNA minor-groove binding agents, such as stapled lexitropins, to enhance phototoxicity. They are not genotoxic. This trait is important in the context of treatment-related secondary malignancies. Conjugation with monoclonal antibodies (e.g., immunoconjugates) affords specificity with respect to the treatment of a variety of diseases, including ovarian cancer and breast cancer. Fluorescence properties of the hypocrellins, or the fact that they are strongly colored, facilitates their use in the diagnosis of tumors and metastatic disease, and their spectral fingerprint facilitates differentiation between malignant, normal, inflamed, or physically damaged tissues. Further, these fluorescence properties can be detected through a variety of optical detection means.

Conjugation with monoclonal antibodies also affords a high degree of phototherapeutic specificity for a variety of diseases, including ovarian and breast cancer. For example, HBEA-R1 transcutaneous phototherapy permanently ablates the EMT6/Ed tumor growing in the flank of Balb/c mice, with minimal cutaneous effects. The conjugates mediate phototoxicity through apoptotic cell death, primarily through type II photochemical reactions with intracellular and membrane targets. In the total absence of oxygen, phototoxicity is mediated through the type I photochemical reaction ("radical cascade"). This feature is critical to the management of hypoxic tumor cells,

1 which currently limit cures by radiation and drugs. They retain significant
phototoxicity at 688 nm, well within the phototherapeutic window [Estey, et al.,
Cancer Chemother. Pharmacol., 37:343 (1996)].

Perylenequinones comprise a growing and highly diverse group of natural
pigments, and they possess some unique chemical and biological properties. The natural
6 perylenequinoid pigments (PQP) identified to date include hypocrellins, cercosporin,
phleichrome, cladochrome, elsinochromes, erythroaphins, and calphostins. Most of
them are produced by a wide variety of molds, and act as the photodynamic
phytotoxins of their hosts, except that erythroaphins are isolated from aphids. For
their general chemical properties [see Weiss, et al., *Prog. Chem. Org. Nat. Prod.*, 52:1
11 (1987) and Diwu, et al., *Photochem & Photobiol.*, 52:609-616 (1990)]. PQP's general
photophysical and photochemical properties have been reviewed in Diwu, et al.,
Pharmac. Ther., 63:1 (1994).

Preliminary data have demonstrated that some PQPs exert a much stronger
photodynamic action on EMT6/ED tumor cells than Photofrin-II®. Hypocrellins,
16 representative of photodynamic therapy applications of PQPs, exhibit several
advantages over the presently used hematoporphyrin derivatives, for example, ready
preparation and easy purification, high triplet quantum yield, strong red light
absorptivity, and significantly reduced normal tissue photosensitivity due to their rapid
excretion. Many of PQP's properties are summarized in Diwu, et al., *J. Photochem.*
21 *Photobiol. A: Chem.*, 64:273 (1992). Some perylenequinones are also potent inhibitors of
certain viruses, particularly human immunodeficiency virus (HIV), and also the enzyme
protein kinase C (PKC). Both anti-HIV and anti-PKC activities of certain PQPs are
light-dependent, a phenomenon implicated in the photodynamic therapy of cancers
[Diwu, et al., *Biochem. Pharmacol.*, 47:373-389 (1994)]. The Diwu et al paper also
26 discloses the successful conjugation of HB to a protein.

As in the case of hematoporphyrin derivatives, natural PQPs do not themselves
exhibit absorptivity longer than 600 nm, a characteristic that inherently describes a
decreased capability of penetrating tissues. This means that the natural PQPs are not
sufficiently strong for photodynamic therapy, and this limits their photodynamic

1 therapy applications.

Deficiencies of current porphyrin photosensitizers for photodynamic therapy have stimulated the development of a series of second generation compounds which have improved properties with respect to light absorption in the red spectral range, purity, pharmacokinetics, and reduced cutaneous photosensitivity. The inventors have concentrated on development of derivatives of the perylenequinone, hypocrellin B. The inventors have identified compounds with excellent physicochemical and biological properties and have completed comprehensive biological characterizations and preclinical evaluation, e.g., Fig. 2, HBEA-R1 (Structure 3) and HBBA-R2 (structure 6).

These compounds, in the presence of 630 or 688 nm light, photopotentiate by a factor of 100-500, with respect to their dark cytotoxicity. HBEA-R1 (Structure 3) and HBBA-R2 (structure 6) have successfully passed dose escalation studies in rodents. No problems of acute or chronic toxicity (60 days or more) were observed following total body doses of 50 μ M. Endpoints considered were body mass, behavior, and gross pathological examination at the end of the observation period. A pharmacokinetic study was completed in Balb/c mice bearing the EMT6/Ed tumor, using 14 C-hypocrellin B (Liu et al., 1995). The information from the HB study provided useful guidelines for timing light administration (Table 1) in the tumor growth delay/control studies with HBEA-R1.

21 Brief Description of the Drawings

Figure 1.: Schematic for the preparation of a variety of aminated Hypocrellin B derivatives. R can be an alkyl, alkenyl, alkynyl or an alcohol containing 2 to 10 carbon atoms.

Figure 2: Chemical structures of hypocrellin A, hypocrellin B, and hypocrellin derivatives, two butylaminated HB structures, two ethanolaminated HB structures, 2-(N,N-dimethylamino)propylamine-HB, and JL-1-1. The phenolic hydroxyl groups of the parent compounds provide a convenient site for modification.

Figure 3: Absorption spectra for HA (dimethylformamide), HB (dimethylformamide), HBBA-R2 (chloroform), HBDP-R1 (chloroform), HBEA-R1

(dimethylsulfoxide), and structure 4 (JL-1-1) (chloroform). Ordinate: Absorbance; Abscissa: Wavelength, nm.

Figure 4. Schematic for the synthesis of radiolabelled hypocrellin B. This compound can then be used to synthesize radiolabelled derivatives of hypocrellin.

Figure 5a-b : Cellular uptake of selected photosensitizers. (a). Standard concentration curves; (b). Uptake by EMT6/Ed cells, adjusted to 10^6 cells. Ordinate: Relative fluorescence at the appropriate detection wavelength (see text); *abscissa*: Concentration of photosensitizer, micromolar.

Figure: 6a-e: Survival curves of cytotoxicity (a) ; (*ordinate* percent survival, *abscissa* photosensitizer concentration, μM) and phototoxicity (b-e); (*ordinate* percentage survival, *abscissa* light dose J/cm^2) of hypocrellin sensitizers, determined by clonogenic assays of EMT6/Ed cells in monolayer culture. b. HBBA-R2, c HBEA-R1, d HBDP-R1, e JL-1-1. Error bars represent standard deviations of five replicate culture plates.

Figure 7. Tumor growth delay under various conditions of HBEA-R1 PDT, of the EMT6/Ed tumor growing in the flanks of Balb/c mice. Transcutaneous illumination (non-hyperthermic).

Figure 8. The effect of light intensity in the for curing tumors in mice in the presence of HBEA-R1. The mice were Balb/c with EMT6/Ed tumours growing in the flanks. The mice were injected with $50 \mu\text{mol}/\text{kg}$ of HBEA-R1 prior to transcutaneous illumination.

Figure 9 a-e. Uptake of (a) hypocrellin B, (b) ^{14}C -HB, (c) HBBA-R2, (d) HBEA-R1 and (e) HBDP-R1 into EMT6/Ed cells versus time.

Figure 10. Oxygen dependency of HBEA-R1 and HBBA-R2 following graded doses of 630 nm light and $0.25 \mu\text{M}$ photosensitizer. Both compounds demonstrate phototoxicity in the absence of oxygen, providing evidence for the type I photochemical process under this condition.

Figure 11. Apoptotic cell death induced as a function of time following PDT treatment of EMT6/Ed cells in monolayer as determined using the propidium iodide method. Sensitizer concentration, $0.20 \mu\text{M}$.

Figure 12. The effect of pH on EMT6/Ed cell cytotoxicity (a) hypocrellin A

1 and 630 nm light, (b) hypocrellin A in the dark, (c) hypocrellin B and 630 nm light, and (d) hypocrellin B in the dark.

Figure 13 shows the synthesis of amino acid derivatives of hypocrellin B.

Figure 14 shows the structure of additional amino acid derivatives of hypocrellin B.

6 Figure 15 shows a scheme for producing hypocrellin B derivatives according to the present invention.

Figure 16 shows a prior art process for producing certain precursor compounds.

Modes For Carrying Out the Invention

11 The present invention comprises the use of perylenequinone (PQP) derivatives as photodynamic agents, and the use of PQP derivatives in photodynamic therapy (PDT). PDT offers a unique treatment alternative for malignancies and the like that are resistant to conventional therapies, with the potential for selective destruction of malignant cells (as shown herein).

16 The invention also comprises a method of treating a disease by administering a therapeutically sufficient amount of at least one PQP derivative, and activating the derivative(s), typically by photoactivating the PQP derivative. Typically, the PQP derivative may be activated by exposing the derivative to a pre-determined wavelength of light. The invention also includes a method of treating cancer which is enhanced in
21 the presence of light wavelengths between about 400 nm and about 850 nm. The absorption spectra for many compounds is shown in Figure 3, and the main absorption peak for each compound is listed in Table 1. Many of these compounds have significant absorbance in the 600 nm to the 700 nm range (see Table 1).

The invention also comprises using one or more PQP derivatives to generate
26 singlet oxygen and a variety of toxic free radicals. Typically, compounds that are capable of generating singlet oxygen and/or toxic free radicals may be used to treat certain diseases and the like.

The invention also comprises using the hypocrellin derivatives that have anti-cancer and/or anti-viral activity, and enhancing the activity of these derivatives by

1 photoactivating the derivative. The invention also includes using the hypocrellin derivatives to preferentially destroy or preferentially target cancer cells.

The invention also comprises a method for producing native perylenequinones, such as hypocrellin, synthesizing perylenequinone derivatives, such as hypocrellin A and hypocrellin B, radiolabeled hypocrellin B and radiolabeled hypocrellin B derivatives. The invention also comprises compositions containing perylenequinone derivatives, hypocrellin derivatives, radiolabeled hypocrellin B and radiolabeled hypocrellin B derivatives, perylenequinone conjugates, and hypocrellin conjugates.

The invention also comprises conjugating the PQP derivatives of the present invention to one or more binding agents, such as antibodies or an antibody fragment.

11 The invention also comprises the PQP derivative conjugated to one or more binding agents, such as an antibody or antibody fragments. The invention also comprises conjugating the PQP derivatives, e.g., hypocrellin derivatives, to DNA minor-groove-binding agents to effect phototoxicity in a cell structure, such as the cell nucleus.

As used herein, "perylenequinone derivative" or "derivative" refers to all compounds derived from native or natural perylenequinones and which can be activated by light of a pre-determined wavelength. In a preferred embodiment of the invention, the derivative is a compound derived from naturally occurring hypocrellin A or hypocrellin B, and hypocrellin-like compounds. Hypocrellin derivatives, as used herein, may be activated by light, and may be used as photodynamic agents. A derivative according to the invention may also be complexed with or include other active reagents, including but not limited to chemotherapeutic agents or alkylating agents. The structures of exemplary derivatives are shown in the Figures.

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As used herein, "disease" refers to the management, diagnosis, and/or palliation of any mammalian (including human) disease, disorder, malady, or condition that can be treated by photodynamic therapy. "Disease" includes but is not limited to cancer and its metastases, such as skin cancer; growths or tumors, and their metastases; tumors and tumor cells, such as sarcomas and carcinomas, including solid tumors, blood-borne tumors, and tumors found in nasal passages, the bladder, the esophagus, or lung, including the bronchi; viruses, including retroviruses; bacterial diseases; fungal diseases;

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1 and dermatological conditions or disorders, such as lesions of the vulva, keloid, vitiligo, psoriasis, benign tumors, endometriosis, Barrett's esophagus, *Tinea capitis*, and lichen amyloidosis.

As used herein, "administering" refers to any action that results in exposing or contacting one or more PQP derivatives with a pre-determined cell, cells, or tissue, typically mammalian. As used herein, administering may be conducted *in vivo*, *in vitro*, or *ex vivo*. For example, a composition may be administered by injection or through an endoscope. Administering also includes the direct application to cells of a composition according to the present invention. For example, during the course of surgery, tumor cells may be exposed. In accordance with an embodiment of the invention, these exposed cells (or tumors) may be exposed directly to a composition of the present invention, e.g., by washing or irrigating the surgical site and/or the cells.

As used herein, "binding agent" refers to any reagent or the like that forms a specific bond with a receptor carried on a target moiety, e.g., an antibody or antibody fragment that binds to the CA 125 antigen of ovarian cancer, or an agent that targets a certain region or structure(s) of a cell. In a preferred embodiment of the invention, the binding agent is an antibody or antibody fragment that specifically binds to cancer cells. In a more preferred embodiment of the invention, the binding agent binds an epitope of an antigen of ovarian cancer, breast cancer, or gastrointestinal cancer (e.g., the CA125 epitope of ovarian cancer, the CA 15.3 epitope of breast cancer, or the CA 19.9 epitope of gastrointestinal cancer).

The antibody or fragment may be labeled (e.g., with radioisotopes or other markers) or unlabeled, and/or in a complex; chimeric monoclonal antibodies ("C-MAb"); genetically engineered monoclonal antibodies ("G-MAb"); fragments of monoclonal antibodies (including but not limited to "F(Ab)₂", "F(Ab)" and "Dab"); single chains representing the reactive portion of monoclonal antibodies ("SC-MAb"); tumor-binding peptides; any of the above joined to a molecule that mediates an effector function; and mimics of any of the above. Various binding agents, antibodies, antigens, and methods for preparing, isolating, and using the antibodies are described in U.S. Patent 4,471,057 (Koprowski) and U.S. Patent 5,075,218 (Jette, et al.), both

1 incorporated herein by reference. Furthermore, many of these antibodies or binding
agents are commercially available from Centocor, Abbott Laboratories, Commissariat à
L'Energie Atomique, Hoffman-LaRoche, Inc., Sorin Biomedica, and FujiRebio.

As used herein, "photopotential factor" refers to the property of the
compound(s) to exert light-mediated toxicity in excess of its (their) inherent dark
6 toxicity. In a preferred embodiment of the invention, the photoactivation factor may be
calculated as the ratio of the LD₅₀ of cells treated without light to the LD₅₀ of the cells
treated with a light-activated compound (drug LD₅₀ divided by drug and light LD₅₀).
Where the term "LD₅₀" has been used above, the term "IC₅₀" may be substituted, to
address the bioassays that concern metabolic activity rather than the endpoint of
11 lethality, loss of reproductive capability, or clonogenic death. The relative
photoactivation efficiency of a compound may also be determined using a clonogenic
assay, an assay that is well known to those skilled in the art (see, for example, Example
13).

In accordance with the present invention, a desirable PQP derivative is one that
16 is non-toxic (or of low toxicity) at high drug concentrations without light (i.e., "dark"),
and is toxic at low concentrations when light of the appropriate wavelength is applied.
As is recognized by those skilled in the art, the most desirable compounds are those that
provide a wide range of non-toxic doses in the dark, as this characteristic provides an
increased safety factor for the patient. As noted in more detail in the Examples, an *in*
21 *vitro* photopotential factor test successfully indicates which hypocrellin congeners
have the best *in vivo* tumor-killing potential. For example, HBEA-R1 and HBBA-R2
photopotential by a factor of 100 or more with respect to their dark cytotoxicities in
the presence of 630 nm or 688 nm light (see Table 1).

As used herein, physiologically acceptable fluid refers to any fluid or additive
26 suitable for combination with a composition containing a PQP derivative. Typically
these fluids are used as a diluent or carrier. Exemplary physiologically acceptable fluids
include but are not limited to preservative solutions, saline solution, an isotonic (about
0.9%) saline solution, or about a 5% albumin solution or suspension. It is intended that
the present invention is not to be limited by the type of physiologically acceptable fluid

1 used. The composition may also include pharmaceutically acceptable carriers.
Pharmaceutically accepted carriers include but are not limited to saline, sterile water,
phosphate buffered saline, and the like. Other buffering agents, dispersing agents, and
inert non-toxic substances suitable for delivery to a patient may be included in the
compositions of the present invention. The compositions may be solutions,
6 suspensions or any appropriate formulation suitable for administration, and are
typically sterile and free of undesirable particulate matter. The compositions may be
sterilized by conventional sterilization techniques.

In accordance with a method of the invention, the binding agent must be capable
of binding a pre-determined binding site or receptor, and may be administered to the
11 patient by any immunologically suitable route. For example, the binding agent may be
introduced into the patient by an intravenous, subcutaneous, intraperitoneal,
intrathecal, intravesical, intradermal, intramuscular, or intralymphatic routes. The
composition may be in solution, tablet, aerosol, or multi-phase formulation forms.
Liposomes, long-circulating liposomes, immunoliposomes, biodegradable
16 microspheres, micelles, or the like may also be used as a carrier, vehicle, or delivery
system. Furthermore, using *ex vivo* procedures well known in the art, blood or serum
from the patient may be removed from the patient; optionally, it may be desirable to
purify the antigen in the patient's blood; the blood or serum may then be mixed with a
composition that includes a binding agent according to the invention; and the treated
21 blood or serum is returned to the patient. The clinician may compare the anti-idiotypic
and anti-isotypic responses associated with these different routes in determining the
most effective route of administration. The invention should not be limited to any
particular method of introducing the binding agent into the patient.

The compounds of the present invention may be produced by any method that
26 results in a purified or substantially purified compound, or in a compound that is useful
as a photodynamic agent. The compounds of the present invention may also form a
composition comprising a cocktail of compounds, e.g., more than one compound.
These methods are well known in the art, e.g., Liu, et al., "Synthetic studies in novel
hypocrellin B derivatives," *Tetrahedron*, 49:10785 (1993); and Diwu, et al., Anti-

1 Cancer Drug Design, 8:129-143 (1993). Hypocrellin derivatives may be readily
synthesized from the parent compound, hypocrellin B (HB), a natural product of the
fungus *Hypocrella bambusae* sacc., a phytopathogen of bamboo. The parent compound
may also be produced synthetically as shown in the Examples. Exemplary methods of
producing the compounds are also shown in more detail in the Examples. It is intended
6 that the invention is not to be limited by the method of producing, isolating, or
purifying the hypocrellin derivatives.

Briefly, for the synthesis of HA, crude HA can be prepared by acetone
extraction of *Hypocrella bambusae* (B. And Br.) Sacc. Lipids can be removed by counter
extraction with petroleum ether. Further purification can be carried out on a silica gel
11 column, followed by 1% potassium dihydrogen phosphate-silica gel thin-layer
chromatography and recrystallization from acetone.

For the synthesis of HB, crude HB can be prepared by quantitative potassium
hydroxide dehydration of HA, followed by neutralization with HCl chloroform
extraction and recrystallization from benzene-petroleum ether. The product was
16 subjected to 1% citric acid-silica gel thin-layer chromatography using a 6:2:1 mixture of
petroleum ether acetate ethanol as eluent. HB derivatives, HBBA-R2 (butylaminated
HB), HBDP-RI (2-(N,N-dimethylamino)-propylamine-HB), and HBEA-RI
(ethanolaminated HB) were prepared by amination of the phenolic hydroxyl groups of
the parent compound. JL-1-1 (structure 4) was prepared according to the method of
21 Liu, et al., *Tetrahedron* 49:10785 (1993). The absorption spectra of these derivatives
were determined on a Hewlett-Packard diode array spectrophotometer (see Figure 3).

Intracellular uptake may be rapid (e.g., within 2 hours for HBEA-R1 and
HBBA-R2), or may uptake may require more time (e.g., about 20 hours for HBDP-R1).
Some degree of selective tumor uptake might be achieved by modification of the pKa of
26 the sensitizer, since the interstitial milieu of some tumors is more acidic than that of
normal tissues. This invention includes a method for identifying compounds where the
toxicity of the compounds is higher for cancer cells than for normal cells, via
comparative clonogenic assays (see Figure 12 and Example 13).

The PQP derivatives of the present invention may also be used in conjunction

1 with and conjugated to a number of other compounds, signaling agents, enhancers,
and/or targeting agents. For example, a hypocrellin derivative of the present invention
may be conjugated to an antibody, preferably a monoclonal antibody. In accordance
with the present invention, the binding agent includes any DNA minor-groove
targeting agent, such as lexitropin or netropsin, preferably to enhance the targeting of
6 the phototoxicity in the cell nucleus. Suitable enhancers include but are not limited to
pKa modifiers, hypoxic cell radiosensitizers, and biochemically activated anti-neoplastic
agents, such as mitomycin C (preferably to effect or potentiate the toxicity of the
compound in hypoxic cells or microorganisms). Suitable signaling agents include but
are not limited to markers of apoptotic cell death or necrotic cell death, or regulatory
11 molecules endogenous to cell cycle control or delay, preferably to potentiate the
phototoxicity of the compound(s) by induction of apoptotic or necrotic cell death, or
by inhibition of the repair of any form of lethal or potentially lethal damage (PLD).

As noted above, an embodiment of the invention includes binding agent-PQP
conjugates (or immunoconjugates) and the therapeutic use of these conjugates. In
16 accordance with the present invention, any method of linking a binding agent to a PQP
may be used. For example, it is well known how to link an antibody or an antibody
fragment to a photosensitizer. For example, Goff, et al., *British Journal of Cancer*,
74:1194-1198 (1996) discloses the production of an immunoconjugate by incubating a
photosensitizer with monoclonal antibody OC125, an antibody that specifically binds
21 to the CA125 antigen associated with most ovarian cancers. In this exemplary
immunoconjugate, polyglutamic acid may be bound to a monoethylendiamine
monoamide derivative, which is then covalently linked to the carbohydrate moiety at
the hinge region of the monoclonal antibody away from the antigen binding sites.
Other exemplary linkages are disclosed in U.S. Patent 4,722,906 and 3,959,078, both
26 incorporated herein by reference. Briefly, these patents disclose providing a
photosensitizer with a selector group, or a latent reactive group, that is the other
member of a specific binding pair, e.g., a reactive group that covalently bonds to an
antibody.

In accordance with the present invention, the PQP derivatives may be

1 functionalized, e.g., include reactive groups including but not limited to an acid, hydroxyl, an acid halide (preferably bromide), a hydrazine, a thiol, or a primary amine. The binding reagent may include reactive groups including but not limited to amino acids, such as cysteine, lysine, aspartic acid, glutamic acid and other dicarboxylic acid amino acids, and other tri- or poly-functional amino acid derivatives.

6 In a preferred embodiment of the invention, the PQP is an amino acid derivative of hypocrellin B, as shown in Figures 13 and 14, and described in Example 20. All R groups are a potential antibody binding site. At the present time, the most preferred immunoconjugates use hypocrellin B derivative 6 in Figure 14, and which include an acid, acid bromide, hydrazine, thiol, or primary amine antibody binding site (e.g., R group). In a preferred embodiment of the invention, any of these derivatives may be linked to a monoclonal antibody that specifically binds to ovarian cancer (e.g., the CA125 epitope), breast cancer (e.g., the CA15.3 epitope), or gastrointestinal cancer (e.g., the CA19.9 epitope). In accordance with the present invention, the most preferred immunoconjugates are hypocrellin B derivative 6 (Figure 14) bound to OC 125, B43, Ar 8.1, or PDL 10 antibodies that specifically bind to ovarian cancer; DF 3 or AR 20.5 antibodies that specifically bind to breast cancer; or B67.4, NS 1116, or AR 18.4 antibodies that specifically bind to gastrointestinal cancer. Many of these, and other, binding agents are commercially available.

21 As is recognized by one skilled in the art, an effective dose of the derivative or a conjugate that includes the derivative will depend in part on the severity of the disease and the status of the patient's immune system. One skilled in the art will recognize that a variety of doses may be used, and are dependent on a variety of well known factors. For example, the dose would be different for diagnosis versus treatment, or for palliation versus management. Generally, the composition will include about 0.1 μ g to about 2 mg or more of binding agent per kilogram of body weight, more commonly dosages of about 200 μ g per kilogram of body weight. The concentration usually will be at least about 0.5%. Any amount may be selected primarily based on fluid volume, viscosity, antigenicity, etc., in accordance with the chosen mode of administration.

Administration of the conjugate or the derivative may be more than once,

1 preferably three times over a prolonged period. As the compositions of this invention
may be used for patients in a serious disease state, i.e., life-threatening or potentially life-
threatening, excesses of the binding agent may be administered if desirable. Actual
methods and protocols for administering pharmaceutical compositions, including
dilution techniques for injections of the present compositions, are well known or will
6 be apparent to one skilled in the art. Some of these methods and protocols are
described in *Remington's Pharmaceutical Science*, Mack Publishing Co. (1982).

In accordance with another embodiment of the invention, a composition of the
present invention may be administered alone, in combination with other compositions,
or in sequence with other PDT compositions. For example, it has been disclosed above
11 that the cellular uptake for HBEA-R1 and HBBA-R2 is rapid and distributed primarily
in cytoplasmic elements. In contrast, it has also been disclosed above that HBDP-R1
reaches peak intracellular concentrations after approximately 20 hours, and is
distributed primarily to cellular membranes. These features afford potential
augmentation of the photodynamic therapeutic ratio through sequential sensitizer
16 administration (followed by light treatment). Under these conditions, a larger number
of organelles can be targeted.

In this embodiment of the invention, a PDT method comprises administering a
first photodynamic agent, preferably having a slow uptake, and administering a second
photodynamic agent, preferably having a more rapid uptake than that of the first agent.
21 Both first and second photodynamic agents may then be activated by exposing the
patient and/or the agent to light of suitable wavelength, as described above.

The excellent fluorescence properties of the hypocrellins and derivatives
provide a valuable tool to monitor intracellular uptake and distribution kinetics by
confocal laser scanning microscopy (CLSM). Each drug has unique properties of uptake
and distribution (Figure 9) (Miller et al 1995 a,b) It is noteworthy that uptake is
26 essentially complete within the first two hours of administration of HBEA-R1
(Structure 3) and HBBA-R2 (structure 6) (Miller et al 1995 a,b). The rate cells take up
drug in humans *in vitro* and *in vivo* can be determined using similar protocols as Liu et
al 1995 and Miller et al., 1995 a or b). *In vivo*, the ideal time between i.v. injection or

1 administration of the drug and light administration is preferably when tumor
concentration of the photodynamic agent is optimal with respect to normal tissues,
typically up to about 24 hours, but as long as 48 hours or more (Table 2).

Some of the embodiments of the present invention also have the added benefit of
functioning with or without the presence of oxygen. Therefore, some embodiments of
6 the present invention are effective in the treatment of solid tumors which are either
well oxygenated or either partially or fully hypoxic.

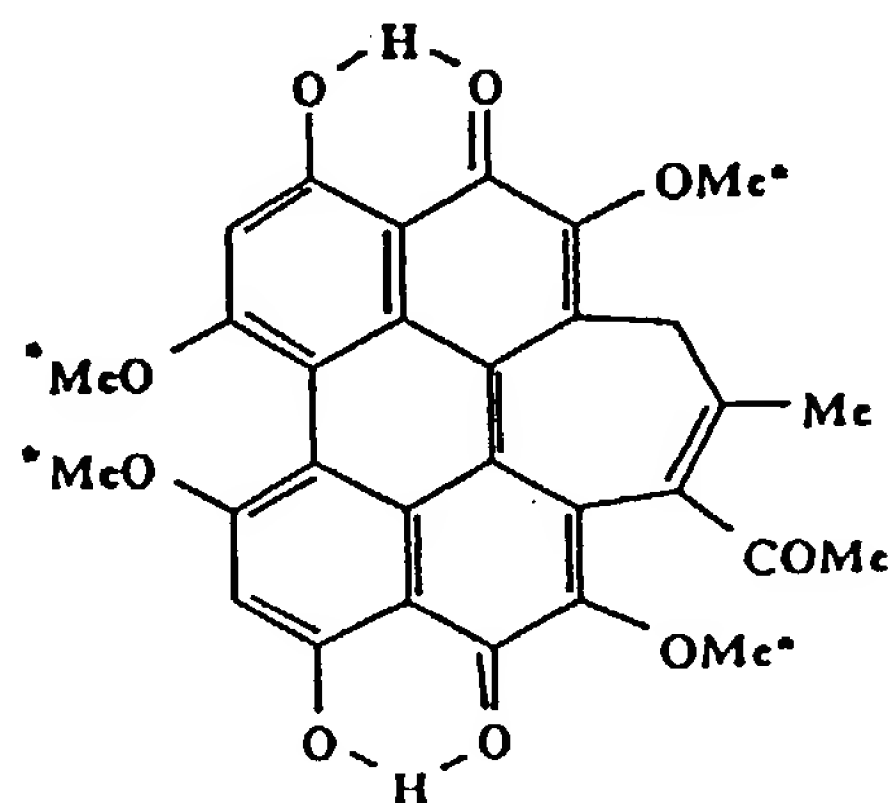
Examples

Example 1. Derivatives obtained from hypocrellin A.

11 Briefly, crude HA was prepared by acetone extraction of *Hypocrella bambusae*
(B. et Br.) Sacc. Lipids were removed by counter extraction with petroleum ether.
Further purification was carried out on a silica gel column, followed by 1% potassium
dihydrogen phosphate-silica gel thin-layer chromatography and recrystallization from
acetone. HB was prepared by quantitative potassium hydroxide dehydration of HA,
16 followed by neutralization with HCl chloroform extraction, and recrystallization from
benzene-petroleum ether. The product was subjected to 1% citric acid-silica gel thin-
layer chromatography, using a 6:2:1 mixture of petroleum ether ethyl acetate ethanol as
eluent. HB derivatives, HBBA-R2, HBDP-R1, and HBEA-R1, were prepared by
amination of the phenolic hydroxyl groups of the parent compound. JL-1-1 (structure
21 4) was prepared according to the method of Liu et al. [1993]. The absorption spectra
of these derivatives were determined on a Hewlett-Packard diode array
spectrophotometer (Figure 3).

1 Example 2. Derivatives obtained from hypocrellin B.

The following discloses a process for synthesizing the following structure:



wherein any or all Me groups (marked with an "*") can be replaced with alkyl groups (branched or straight chain), with 2 to 6 carbons, or other chemical groups. These compounds and processes can be used to make precursors or intermediates for compounds including but not limited to anti-cancer agents, anti-viral agents, anti-retroviral agents, anti-bacteriocidal agents, anti-fungal agents, perylenequinones and derivatives, hypocrellins and derivatives, hypocrellin A and derivatives, hypocrellin B and derivatives, cercosporin and derivatives, phleichrome and derivatives, elsinochromes and derivatives, cladochromes and derivatives, erythroaphins and derivatives, calphostins and derivatives, , and other compounds with photodynamic activity.

An efficient total synthesis of HB using precursor 6 is shown in Figure 15. Commercially available 3,5-dimethoxybenzaldehyde was converted to compound 5 using seven reaction steps, according to the procedure shown in Hauser, et al, *J. Org. Chem.* 59:1967 (1994). Oxidation of compound 5 with benzeneseleninic anhydride in anhydrous THF produced the 1,2-naphthoquinone 6 in 87% yield. Attempted coupling of compound 6 to the corresponding perylenequinone by TFA and FeCl_3 was not successful, but coupling of its ester, which was prepared in the presence of Ac_2O ,

1 pyridine, and a catalytic amount of DMAP in 95% yield, gave perylenequinone 7 in
88% yield. Methylation of 7 with methyl iodide and C_5F_5 in anhydrous THF led to
compound 8 quantitatively. Hydrolysis of 8 in K_2CO_3 and MeOH- H_2O solution,
followed by oxidation with CrO_3 and pyridine in CH_2Cl_2 gave the methyl ketone 1 in
63% overall yield. Compound 9, which was obtained from 1 in 85% yield by an
6 intramolecular aldol condensation reaction, was demethylated selectively with 48%
hydrobromic acid to afford a 1:1 mixture of enantiomers HB (90%) which was identical
in all respects (TLC, IR, NMR) with the natural product HB. This synthesis provides
an entry to the synthesis of other perylenequinones.

11 1-acetyloxy-3-acetonyl-6,8-dimethoxynaphthalene (3): this compound was prepared
using six reaction steps from 3,5-dimethoxybenzaldehyde according to the procedures
disclosed in Hauser, et al, *J. Org. Chem.* 59:1967 (1994).

16 1-hydroxy-3-acetonyl-6,8-dimethoxynaphthalene (4): to a solution of 3 (624 mg., 2.07
mmol) in MeOH (20 ml) was added K_2CO_3 (1.0 g) in water. (10 ml), and the mixture
was stirred for two hours, acidified with 5% HCl, diluted with water (100 ml), and
extracted with $CHCl_3$. The combined extracts were washed with water, then dried
(Na_2SO_4), filtered and evaporated to give product 4 (505 mg, 94% yield). 1H NMR: δ
9.10 (s, 1H, ArOH), 7.00 (d, 1H, $J = 2.0$ Hz, ArH), 6.68 (d, 1H, $J = 2.0$ Hz, ArH), 6.57
21 (d, 1H, $J = 2.0$ Hz, ArH), 6.40 (d, 1H, $J = 2.0$ Hz, ArH), , 4.00 (s, 3H, OMe), 3.88 (s,
3H, OMe), 3.70 (s, 2H, CH_2), 2.18 (s, #H, Me). HRMS (m/e) $C_{15}H_{16}O_4$: (M) calc.
260.10486; found 260.10472.

(\pm) -4-hydroxy-5,7-dimethoxy- α -methylnaphthalene-ethanol (5): this compound was
prepared according to the procedures disclosed in Hauser, et al, *J. Org. Chem.* 59:1967
26 (1994).

(\pm) -3-[2-(hydroxy)propyl]-6,8-dimethoxy-1,2-naphthaquinone (6): a solution of 5
(440 mg, 1.53 mmol) in dry THF (15 ml) was added dropwise during 15 minutes to a
stirred suspension of 70% benzeneseleninic anhydride (900 mg, 1.75 mmol) in THF (15
ml) at 50°C and stirring was continued for 10 minutes. The reaction mixture was

1 poured into water (100 ml) and extracted with CHCl_3 . The extract was washed with
aqueous NaHCO_3 solution (10%) and water, and then dried. Evaporation followed by
column chromatography on silica gel using CH_2Cl_2 :MeOH (20:1 v/v) as an eluent gave
the product 6 (402 mg, 87% yield) as an orange solid, m.p. 70-72°C. ^1H NMR: δ 7.12
(s, 1H, ArH), 6.38 (d, 1H, $J = 2.0$ Hz, ArH), 6.36 (d, 1H, $J = 2.0$ Hz, ArH), 4.04 (m,
6 1H, CHOH), 3.91 (s, 3H, OMe), 3.89 (s, 3H, OMe), 2.67 (m, 1H, CHaHb), 2.62 (m,
1H, CH'aH'b), 2.41 (m, 1H, CHaHb), 2.36 (m, 1H, CH'aH'b), 1.25
(s, 3H, Me) 1.24 (s, 3H, Me'). HRMS (m/e) $\text{C}_{15}\text{H}_{16}\text{O}_4$: (M) calc. 276.09976; found
276.09882.

11 3-Acetyl-6,8-dimethoxy-1,2-naphthoquinone (2):

This compound was prepared in 81% yield from 4 in a similar manner as that
described for 6. This compound is unstable and can not be purified by
chromatography, but it can be recrystallized from CHCl_3 , melting point 185°C (dec.).
 ^1H NMR: δ 7.19 (s, 1H, ArH), 6.46 (d, 1H, $J = 2.0$ Hz, ArH), 6.44 (d, 1H, $J = 2.0$ Hz,
16 ArH), 3.97 (s, 3H, OMe), 3.92 (s, 3H, OMe), 3.55 (s, 2H, CH_2), 2.29 (s, 3H, Me).
HRMS (m/e) $\text{C}_{15}\text{H}_{16}\text{O}_5$: (M + 2H) calc. 276.09976; found 276.09946.

To a solution of 6 (300 mg, 1.09 mmol) in CHCl_3 (10 ml) was added Ac_2O (1.5
ml), pyridine (1.5 ml) and 4-dimethylaminopyridine (DMAP) (100 mg). The reaction
mixture was stirred for 3 hours at room temperature, diluted with H_2O (100 ml), and
21 extracted with CHCl_3 . The extracts were washed with 1N HCl and H_2O respectively.
The residue was purified by flash chromatography on silica gel using CH_2Cl_2 :MeOH
(50:1, v/v) as an eluent to give the ester (330 mg, 95% yield), melting point 62-64°C. ^1H
NMR: δ 7.07 (s, 1H, ArH), 6.42 (d, 1H, $J = 2.0$ Hz, ArH), 6.40 (d, 1H, $J = 2.0$ Hz,
ArH), 5.09 (m, 1H, CHOAc), 3.96 (s, 3H, OMe), 3.92 (s, 3H, OMe), 2.76 (m, 1H,
26 CHaHb), 2.72 (m, 1H, CH'aH'b), 2.61 (m, 1H, CHaHb), 2.56 (m, 1H, CH'aH'b), 1.59
(s, 3H, Oac), 1.27 (s, 3H, Me), 1.25 (s, 3H, Me'). HRMS (m/e) $\text{C}_{17}\text{H}_{18}\text{O}_6$: (M) calc.
318.11035; found 318.10938.

1 **1,12-Bis[2-(acetyloxy)propyl]-4,6,7,9-tetramethoxy-2,11-dihydroxy-3,10-
perylene-1,9-dione (7):**

 A solution of the naphthoquinone (200 mg, 0.63 mmol) in CHCl_3 (4.0 ml) and TFA (1.5 ml) was stirred at room temperature under N_2 for 20 minutes. A solution of anhydrous FeCl_3 (51 mg, 0.31 mmol) in CH_3CN (5.0 ml) was slowly added dropwise
6 over one hour. The reaction, monitored by TLC, indicated no starting material remained. The resulting mixture was diluted with H_2O (100 ml) and extracted with CHCl_3 . The extracts were washed with H_2O , dried (Na_2SO_4), filtered and evaporated. Chromatography of the residue on silica gel using CHCl_3 :MeOH (50:1 v/v) as an eluent gave the product (175 mg, 88% yield). ^1H NMR: δ 8.09 (s, ArH), 8.01 (s, ArH), 6.76
11 (m, ArH), 4.83 (m, CHOAc), 4.24–4.12 (m, OMe), 3.70 – 2.86 (m, CH_2), 1.75 (s, OAc), 1.02 (m, Me), 0.90 (s, OAc), 0.88 (s, OAc), 0.59 (m, Me). HRMS (m/e) $\text{C}_{34}\text{H}_{36}\text{O}_{12}$: (M + 2H) calc. 636.22070; found 636.22067.

1,12-Bis[2-(acetyloxy)propyl]-2,4,6,7,9,11-hexamethoxy-3,10-perylene-1,9-dione (8) :

16 To a solution of 7 (250 mg, 0.39 mmol) in anhydrous THF (10 ml) was added CsF (500 mg) and methyl iodide (0.5 ml). After stirring overnight at room temperature, the reaction mixture was diluted with H_2O (100 ml) and extracted with CHCl_3 . The combined extracts were dried (Na_2SO_4), filtered and evaporated. Chromatography of the residue on silica gel using CHCl_3 :MeOH (60:1, v/v) as an
21 eluent gave the product (245 mg, 94% yield). ^1H NMR: δ 6.77 (m, ArH), 4.73 (m, CHOAc), 4.63 (m, CHOAc), 4.19–4.08 (m, OMe), 3.43 – 2.62 (m, CH_2), 1.81 (s, OAc), 1.79 (s, OAc), 1.05 (m, Me), 0.76 (s, OAc), 0.75 (s, OAc), 0.52 (m, Me). HRMS (m/e) $\text{C}_{36}\text{H}_{38}\text{O}_{12}$: (M) calc. 662.23633; found 662.23610.

26 **1,12-Bis[2-(hydroxy)propyl]-2,4,6,7,9,11-hexamethoxy-3,10-perylene-1,9-dione:**

 To a solution of 8 (160 mg, 0.24 mmol) in MeOH (10 ml) was added K_2CO_3 (500 mg) in H_2O (5 ml) and the mixture was stirred overnight, acidified with 5% HCl, diluted with H_2O (100 ml), and extracted with CHCl_3 . The combined extracts were washed with H_2O , then dried (Na_2SO_4), filtered and evaporated to give product (134

1 mg, 96% yield). ^1H NMR: δ 6.76 (m, ArH), 4.19 – 4.07 (m, OMe), 3.43 – 2.64 (m, CH + CH_2), 0.91 (m, Me), 0.68 (m, Me). HRMS (m/e) $\text{C}_{32}\text{H}_{34}\text{O}_{10}$: (M) calc. 578.21521; found 578.21388.

1,12-Bisacetonyl-2,4,6,7,9,11-hexamethoxy-3,10-perylenedione (1):

6 To a solution of dry pyridine (300 μl) in CH_2Cl_2 (5.0 ml) was added CrO_3 (300 mg), and then the mixture was stirred 15 minutes. 40 mg (0.069 mmol) of above compound in CH_2Cl_2 (2.0 ml) was added to the solution and stirred rapidly for 1 minute. The resulting solution was washed with H_2O , dried (Na_2SO_4), filtered and evaporated. Chromatography of the residue on silica gel using CH_2Cl_2 :MeOH (50:1, 11 v/v) as an eluent gave the product (26 mg, 66% yield), melting point 171 – 173 °C. ^1H NMR: δ 6.74 (s, 2H, 2 x ArH), 4.16 (d of AB quartet, $J = 16.0$ Hz, 2H, 2 x CH_{2a}), 4.07 (s, 6H, 2 x OMe), 3.95 (s, 6H, 2 x OMe), 3.50 (d of AB quartet, $J = 16.0$ Hz, 2H, 2 x CH_{2b}). HRMS (m/e) $\text{C}_{32}\text{H}_{30}\text{O}_{10}$: (M) calc. 574.18390; found 574.18126.

16 3-Acetyl-4,6,8,9,11,13-hexamethoxy-2-methyl-1H-cyclohepta[ghi]perylene-5,12-dione (9):

To a solution of 1 (20 mg, 0.035 mmol) in MeOH (3.0 ml) was added LiOH (100 mg) in H_2O (1.0 ml) at 0 °C. The reaction mixture was stirred for 40 minutes at room temperature, diluted with H_2O (100 ml), acidified with 1N HCl, and extracted 21 with CHCl_3 . The combined extracts were dried (Na_2SO_4), filtered and evaporated. Chromatography of the residue on silica gel using CHCl_3 :MeOH (15:1, v/v) as an eluent gave the product (16.5 mg, 85% yield), melting point 154 – 156 °C. ^1H NMR: δ 6.83 (s, 1H, ArH), 6.82 (s, 1H, ArH), 4.21 (s, 3H, OMe), 4.20 (s, 3H, OMe), 4.12 (s, 6H, 2 x OMe), 4.08 (s, 3H, OMe), 4.05 (s, 3H, OMe), 3.77 (d of AB quartet, $J = 11.0$ 26 Hz, 1H, CH_{2a}), 3.26 (d of AB quartet, $J = 11.0$ Hz, 1H, CH_{2b}), 2.28 (s, 3H, Me), 1.99 (s, 3H, Me). HRMS (m/e) $\text{C}_{32}\text{H}_{28}\text{O}_9$: (M) calc. 556.17334; found 556.17437.

Hypocrellin B (HB):

To a solution of 9 (15 mg, 0.027 mmol) in CHCl_3 (2 ml) was added 1.0 ml of

1 48% HBr with stirring for one hour at 25°C. The mixture was poured into water (10 ml), extracted with CHCl₃ and dried. The residue was purified by preparative TLC using CHCl₃ as the developing solvent to afford a 1:1 mixture of enantiomers HB (13.0 mg, 90% yield) which was identical in all respects (TLC, IR, and NMR) with the natural product HB.

6 Another synthetic approach to the production of HB derivatives based on others' disclosed processes is shown in Figure 16. Briefly, simplification of HB by retro-aldol condensation of methyl ketone provides compound 1. Cleavage of the two biaryl bonds of 1 in a retrosynthetic sense affords 3-acetyl-6,8-dimethoxy-1,2-naphthoquinone (compound 2). Takuwa, et al., *Synthesis*, 315 (1993) disclosed a one-pot
11 photochemical reaction to prepare 3-(2-oxoalkyl)-1,2-naphthoquinone, however, the process afforded only a very low yield of the precursor 2 since the two methoxy substituents in the naphthoquinone may markedly decrease the photoreactivity. Hauser, et al, *J. Org. Chem.* 59:1967 (1994) disclosed the total synthesis of calphostin D, and provided a method to prepare precursor 3 in high overall yield. Hydrolysis of 3
16 followed by oxidation affords 2 in high yield. The precursor 2 is unstable and was used immediately for the next coupling reaction by FeCl₃ or TFA but no anticipated product was obtained.

Example 3. Direct amination of hypocrellin B.

21 HB (50 mg) was dissolved in ethanol (5 mL) containing the amine (1 mL), and the resulting solution was refluxed for 6-18 h depending upon the individual amine used. The mixture was poured into ice-water, neutralized with 10% hydrochloric acid, and extracted with chloroform. The chloroform layer was washed with water, and dried with anhydrous Na₂SO₄ and evaporated to afford a blue solid. The solid was first
26 chromatographed on a 1% KH₂PO₄-silica gel column with dichloromethane-methanol (gradient ratio) as an eluent to give several constituents. Each constituent was twice rechromatographed on 1% citric acid-silica gel plate using 6:1:1 petroleum ether-ethyl acetate-ethanol as developing agent to afford the individual derivatives.

1 Example 4. Amination of hypocrellin B with ethanolamine.

Reaction of HB with ethanolamine according to the above procedure affords five products. Compound 8 (Structure 8 in Figure 2 also referred to as 3A or HBEA-R2) and compound 3 ((Structure 3 in Figure 2 also referred to as HBEA-R1 or 3B) (Isomer B)) (Diwu et al. 1993) were identified and characterized as follows:

6 Structure 8 (Figure 2) (20%): R: 3270, 1717 and 1612 cm^{-1} ; ^1H -NMR (in DMSO-d_6): 11.46 (s, < 1H, exchangeable with D_2O , phenolic OH), 1.38 (s, < 1H, exchangeable with D_2O , phenolic OH), 6.83 (s, 1H, ArH), 6.78 (s, 1H, ArH), 4.09 (s, 3H, OCH_3), 3H, OCH_3), 3.94 (s, 3H, OCH_3), 3.92 (s, 3H, OCH_3), 3.85- 3.50 (m, 4H, 2NHCH_3), 3.40- 2.92 (m, 4H, CH_2OH), 2.11 (s, 3H, COCH_3) and 1.72 ppm (s, 3H, CH_3). MS (FAB):
11 615 (M+H). Calculated for $\text{C}_{34}\text{H}_{34}\text{N}_2\text{O}_9$: 614.2264; found, 614.2270.

Structure 3 in Figure 2 [also referred to as 3B or HBEA-R1 (Isomer B)] (12%):
IR: 3260, 1720 and 1613 cm^{-1} ; ^1H -NMR (in DMSO-d_6): 12.11 (s, < 1H, exchangeable with D_2O , phenolic OH), 11.99 (s, < 1H, exchangeable with D_2O , phenolic OH), 6.47 (s, 1H, ArH), 6.35 (s, 1H, ArH), 4.03 (s, 3H, OCH_3), 3.95 (s, 6H, $2 \times \text{OCH}_3$), 3.93 (s,
16 3H, OCH_3), 3.88-3.62 (m, 4H, 2NHCH_3), 3.20-2.95 (m, $2\text{CH}_2\text{OH}$), 2.15 (s, 3H, COCH_3) and 1.90 ppm (s, 3H, CH_3). MS (FAB): 615 (M+H). Calculated for $\text{C}_{34}\text{H}_{34}\text{N}_2\text{O}_9$: 614.2264; found; 614.2268.

Example 5. Amination of hypocrellin B with butylamine.

21 Synthesis of structure 6 (Figure 2 structure 6 also referred to as HBBA-R2 or 4A) (Isomer A) and structure 9 (Figure 2 structure 9 also referred to as 4B) (Diwu et al. 1993). Reaction of HB with butylamine according to i.e. above procedure afforded five products. Two of these compounds were identified as follows:

HBBA-R2 (Structure 6 in Figure 2) (21%): IR: 3280, 1702 and 1616 cm^{-1} ; ^1H -
26 NMR: 15.65 (s, 1H, exchangeable with D_2O , phenolic OH), 14.94 (s, 1H, exchangeable with D_2O , phenolic OH), 6.41 (s, 1H, ArH), 6.40 (s, 1H, ArH), 4.07 (s, 3H, OCH_3), 4.00 (s, 3H, OCH_3), 3.96 (s, 3H, OCH_3), 3.93 (d, 3H, OCH_3), 3.24 (m, 4H, 2NHCH_2), 1.98 (s, 3H, COCH_3), 1.26 (s, 3H, CH_3) and 1.70- 0.85 ppm (m, 14H, $2\text{CH}_2\text{CH}_2\text{CH}_3$). MS (FAB): 639 (M+H). Calculated for $\text{C}_{38}\text{H}_{42}\text{N}_2\text{O}_7$: 638.2992; found; 638.2998.

1 Compound 4B or HBBA-R1 (structure 9 in Figure 2) (11%): IR: 3300, 1715 and
1616 cm^{-1} , ^1H -NMR: 15.40 (s, 1H, exchangeable with D_2O , phenolic OH), 15.18 (s, 1H,
exchangeable with D_2O , phenolic OH), 6.48 (s, 1H, ArH), 6.33 (s, 1H, ArH), 4.01 (s,
6H, 2 x OCH_3), 3.97 (d, 1H, CH), 3.96 (s, 6H), 2 x OCH_3), 3.54 (m, 4H, 2NHCH₂),
3.14 (d, 1H, CH), 2.16 (s, 3H, COCH_3), 1.69 (s, 3H, CH_3) and 1.60-0.85 ppm (m, 14H,
6 2CH₂CH₂CH₂). MS (FAB): 639 (M+H). Calculated for $\text{C}_{38}\text{H}_{42}\text{N}_2\text{O}_7$: 639.2998; found;
638.2992.

Example 6. Synthesis of JL-1-1 (structure 4 in Figure 2)

JL-1-1 was produced according to the process(es) disclosed in Liu, et al.,
11 *Tetrahedron* 49:10785 (1993).

Example 7. Synthesis of HBDP-R1 (structure 7 in Figure 2)

HBDP-R1 was produced according to the process(es) disclosed in Diwu, et al., *J.*
Photochem. Photobiol. B: Biol. 18:131 (1993).

Example 8. Radiolabeled derivatives.

The invention provides a method for preparing radiolabelled HB and
derivatives. The method for synthesizing ^{14}C -HB is described in Liu et al (1995). HB
was dissolved in 6.0 mL of anhydrous benzene containing 130 mg of anhydrous
21 aluminum chloride, and the solution was refluxed for 1 hr under nitrogen. The
mixture was poured into 10% aqueous ammonium fluoride, and extracted with
chloroform. The chloroform layer was washed with water, dried (Na_2SO_4) and
evaporated to afford a red solid that contained compounds 2,3 and 4 as illustrated in
Figure 4 (Diwu and Lown 1993b). A schematic for the synthesis of radiolabeled
26 Hypocrellin B is illustrated in Figure 4.

The red solid was dissolved in 2.0 ml of anhydrous tetrahydrofuran (THF)
containing 200 mg of cesium fluoride, and then ^{14}C -methyl iodide (1.0 mCi) in 1.0 ml of
anhydrous THF was injected into the sealed flask. The mixture was stirred at room
temperature for 24 hr, and stirring was continued another 12 hr at room temperature

1 following the addition of 0.3 ml of methyl iodide. The resulting solution was added to
water and extracted with chloroform. The organic layer that contained compounds 1, 5
, and 6 illustrated in Figure 4 was concentrated to about 5.0 ml by nitrogen stream
evaporation.

6 A 48% hydrobromic acid solution (2.0ml) was added to the chloroform solution
and the mixture was stirred at room temperature for 30 minutes. The reaction mixture
was poured into water, and extracted with chloroform. Evaporation of the chloroform
layer gave a red solid. The solid was chromatographed on silica gel using chloroform as
an eluent to afford radiolabeled HB.

11 This compound can then be used to synthesize radiolabeled derivatives of
hypocrellin B as per the procedures described herein.

These radiolabeled compounds can be used to study drug distribution in tissues
and can be utilized to develop or verify drug delivery systems.

Example 9. Other aminated products

16 Other aminated compounds with R= to an alkyl, alkenyl, alkynyl or an alcohol
containing 2 to 10 carbons (Figure 1) can be readily synthesized based on procedures
described herein. Those with ordinary skill can readily ascertain or determine the
appropriate solvents, other appropriate reagents and conditions without resorting to
undue experimentation.

21 Singlet oxygen yields were determined by the 9,10 diphenylanthracene (DPA)
bleaching method [Diwu and Lown 1992]. Test sensitizers were illuminated at a
predetermined wavelength and the kinetics of decrease in the DPA absorption peak of
374 nm were followed as $^1\text{O}_2$ was accepted by DPA. The physical and chemical
properties of hypocrellin compounds synthesized and screened for toxicity *in vitro* are
26 listed in Table 1. Compounds with exceptionally favorable characteristics of dark
cytotoxicity and photopotential are boldfaced in Table 1.

The molecular weight of the hypocrellins and their derivatives averages
approximately 640 Daltons (range, 528 - 780 D). For perspective, this contrasts with a
larger molecular weight of approximately 1,130 - 4,520 D for the mainly ester- and

1 ether-linked oligomers of hematoporphyrin (2 - 8 porphyrin units) thought to comprise the bulk of Photofrin®.

The hypocrellins reported in this study have absorption bands in the red spectral region (Figure 3). Compounds with significant absorption around 630 nm are highlighted with asterisks in the Absorbance Peak column of Table 1. Absorption spectra of the parent compounds, HA and HB, and of four efficient congeners that are photosensitizers, are illustrated in Figure 3. It is useful to note that singlet oxygen yield (Table 1) is not strongly correlated with photopotential.

Example 10. Tumor cure

11 Using the Balb/c mouse/EMT6/Ed tumor model, the inventors have demonstrated tumor cures using HBEA-R1 (Structure 3) and clinically relevant transcutaneous doses of 630 nm light (100 mW fluence rate). These results are summarized in Figure 8. The conditions of light treatment did not provide a hyperthermic effect, and a light dose of 100 J/cm² effected 80 day cures in 9 of 10 animals tested to date. In fact, no animal exhibited any evidence of tumor re-growth, even until the time of sacrifice (100 days post-treatment). At the constant sensitizer "whole body" dose of 50 µmol/kg, reduction of the light dose provided less efficient tumor control. Light delivery approximately 2 hours following HBEA-R1 (Structure 3) administration provides good tumor control in the absence of marked cutaneous phototoxicity. Following a period of dry erythema and subsequent eschar formation, skin overlying the treated tumors returned to complete cosmetic normalcy within 30 days. It is of interest that HBEA-R1 PDT (≥ 100 J/cm² 630) does not evoke *any* skin response in tumor-free Balb/c mice treated with 50 µmol/kg HBEA-R1 at graded intervals to 48 hours. These results provide encouragement that HBEA-R1 will defeat the major limitations of prolonged cutaneous photosensitivity exerted by Photofin®.

Due to the potential for regional hypoxia in some human tumors and the requirement for oxygen for the photodynamic effect, it is important to understand the relative contributions of type I and type II photochemical reactions to phototoxicity. Findings in EMT6/Ed cells in suspension culture with stringently controlled oxygen

1 concentrations in the gas phase during PDT, have revealed that the type II reaction predominates. Nevertheless, there is a component of type I phototoxicity when cells are sensitized with HBEA-R1 or HBBA-R2 (structure 6) in the absence of oxygen.

6 Example 11. Cellular Uptake of Photosensitizers

Standard curves of fluorescence yield vs. concentration were prepared for selected compounds dissolved in 3.6 mL dimethylsulfoxide (DMSO) mixed with unlabeled EMT6/Ed cells which had been removed from Petri dishes with 0.4 mL of 0.5% Tween 20 in 1 N NaOH. This procedure resulted in equivalence of the standard
11 curves with the actual cell uptake assays. Cells labeled for two hours with hypocrellin B and selected congeners were rinsed three times with phosphate-buffered saline to completely remove excess drug, removed from Petri plates as described above, and added to DMSO. The washing procedure was monitored spectrophotometrically. The Tween 20-NaOH solution used to remove the cells from the plates resulted in complete
16 cytolysis and extraction of the hypocrellins from the cells. Fluorescence was quantified with a Spex Fluoro Max™ fluorometer. The illumination and detection slit widths were set at 1 nm. The optimal excitation wavelengths were determined for each compound. For example HB excitation was 438 nm, HBBA-R1, 411 nm; HBEA-R1, 352 nm; HBDP-R1, 410 nm; and JL-1-1, 594 nm. Uptake per 10⁶ cells was determined
21 by regression analysis of the individual uptake curves, and is outlined below. A standard, 2-hour incubation interval was chosen to match the empirically chosen cytotoxicity/phototoxicity pre-incubation. Intracellular uptake kinetics experiments performed by fluorescence confocal microscopy demonstrate that intracellular uptake is complete within the two hour incubation interval (Figure 9) (Miller, et al. 1995 a,b).

26 Example 12. Cell Line and Culture Conditions

EMT6/Ed mouse tumor cells have been propagated by the inventors for several years [Chapman et al. 1983]. They are maintained as monolayer cultures in Waymouth's medium containing 12.5% fetal calf serum, at 37°C in a humidified

1 atmosphere of 95% air and 5% CO₂. They require twice weekly transfers to maintain
their exponential growth. A minimum of three times per year, the cell line is passaged
as solid tumors in BALB/c mice. Newly passaged EMT6/Ed cell lines are reestablished
and propagated for at least two weeks *in vitro* prior to experimentation. This
procedure maintains the malignant phenotype.

6

Example 13. Photosensitizers and Clonogenic Assays

Purified hypocrellin derivatives were obtained in lyophilized form. One day
prior to each experiment, exponentially growing EMT6/Ed cells (25,000/mL in 2.0 mL)
were seeded in 3.0 cm tissue culture Petri plates. The photosensitizers were kept in
11 lyophilized form until the day of experiment, at which time they were dissolved in
DMSO. Stock sensitizer-DMSO solutions were diluted with Hank's Balanced Salts
Solution and added to the cells in Waymouth's medium containing 12.5% fetal calf
serum for the sensitization studies. The maximum concentration of DMSO in the
incubation medium was 1% (v/v), and any effects of DMSO on cloning efficiency were
16 controlled by series of dishes containing the appropriate concentration of DMSO, but
no sensitizer. Preliminary studies indicate that the presence of fetal calf serum did not
significantly influence toxicity. EMT6/Ed cells were exposed to graded doses of the test
compound for 2 h, after which time the test compound was removed by repeated
washing with Hank's Balanced Salts Solution. The cells were then illuminated as
21 monolayers with graded doses of 630 nm light in room air (*vide infra*). The cells were
trypsinized, counted, and plated at known density in Waymouth's media. The cultures
were incubated for 6 days and subjected to standard clonogenic assay [Puck and Marcus,
1956].

The dark toxicity characteristic of each compound was assessed separately
26 following a similar procedure for cell exposure to graded doses of the photosensitizer
for 2 h. Precautions were taken to avoid exposure of the cells to light throughout the
period that cells were exposed to, or contained, photosensitizer. Survival curves
represent a minimum of three independent experiments. Data were corrected for
plating efficiency and the phototoxicity curves were corrected for dark toxicity for each

1 drug concentration, where necessary.

Example 14. Illumination of the Cell Cultures

6 A 20 Watt argon tunable dye laser system (Coherent, Innova-20, Model CR599) with Kiton Red dye (Exciton, Dayton, Ohio) was used to generate light at 630nm. The wavelength was verified with a monochromator prior to experimentation. The light beam was transported by a single, 0.4 mm quartz optical fibre and oriented 9.3 cm. above the cell culture dish. The terminus of the fibre was fitted with a lens which evenly illuminated the flat surface of the irradiation table. Rotation of the table during illumination optimized uniformity of light distribution. The dose at the surface of the cell culture was determined by a power meter (Coherent - 210) to be 1 J/ cm². Illuminations were performed at ambient temperature (~ 23°C) and the longest exposure time was 1 min.

16 The sensitizer uptake studies were performed under incubation conditions identical to those used for clonogenic assays. Figures 5 a. and b. demonstrate the standard curves and cellular uptake data, respectively. The molar quantity of sensitizer required to effect 50% lethality per 10⁶ cells in the dark was estimated by inspection of the dark toxicity survival curves Figure 6 a., (*vide infra*) and observation of intracellular concentration achieved for the appropriate extracellular drug concentration from Figure 5b. For HB, this value is 8 nanomoles; HBEA-R1, 120 nanomoles; and JL-1-1, 32 nanomoles. Application of 630 nm light markedly reduced intracellular photosensitizer quantities required to achieve 50% cell killing. For light doses up to J/ cm², 160 pmole/10⁶ cells of HB, or a 50-fold reduction in bound drug is required. HBEA-R1 is effective at 1 nmole/10⁶ cells (120-fold less); HBDP-R1, 11 nmole/10⁶ cells; and JL-1-1, 0.32 nmole/10⁶ cells, a 100-fold photopotential of 50% lethality. Among the compounds tested, the concentration range for LD₅₀ in the clonogenic assay ranged from approximately 10 μM to > 100 μM (mean, approx. 25 μM). In the presence of 630 nm light, the LD₅₀ photosensitizing dose of hypocrellins was reduced (0.15 to > 6 μM mean, approx. 3 μM), in some cases, substantially. HBBA-R2 demonstrated up to 500 - fold photopotential *in vitro*, while HBEA-R1, HBDP-R1 (Figure 2 structure 7)

1 and JL-1-1 were characterized by 167-, 50-, and 35-fold photopotential factors,
respectively. The photopotential values based on clonogenic assays generally vary
proportionately with drug uptake, for those compounds for which both values were
determined.

Survival curves of the *in vitro* dark toxicity of four hypocrellin derivatives are
6 displayed in Figure 6a. Error bars represent the standard deviations of 5 replicate
culture plates from three independent experiments for each compound. Each derivative
has a characteristic cytotoxicity. HBBA-R2 and JL-1-1 did not evoke 50% lethality
within the concentration range tested ($\leq 80 \mu\text{M}$). HBDP-R1 produced 50% cell death at
concentrations in excess of $40 \mu\text{M}$. HBEA-R1 is effective in the $20 \mu\text{M}$ range. Many of
11 the compounds in table 1 are photopotential and the LD_{50} for many of these
compounds are lower than HA or HB.

Some cancer cells are slightly more acidic than normal cells therefore
compounds that are more active at a pH of 6.5 than 7.0 are desirable for treating some
cancers. HB (Figure 12c and d) and some of the other derivatives are most active at pH
16 6.5 (± 0.5) and should be more active in acidic cells. The ability of HA to kill
EMT6/Ed cells, on the other hand, does not appear to be affected by changes in pH
from 5.5 to 7.5 (Figure 12a and b).

The excellent photopotential characteristic of HBBA-R2 is due to extremely
low dark cytotoxicity on the one hand, and excellent photo-sensitization, on the other
21 (Figures. 6a and b.). HBEA-R1 is another excellent photosensitizer, with a reciprocal
drug-light dose response in the $0.15\text{-}0.3 \mu\text{M}$, $0.25\text{-}1.00 \text{ J/cm}^2$ ranges (Figures 6a and c.).
Data presented in Figure 6d. indicate that the LD_{50} or phototoxicity of HBDP-R1 lies in
the $0.5\text{-}2.5 \mu\text{M}$ range, in the presence of 0.75 J/cm^2 , 630 nm light. Finally, the toxicity
of JL-1-1 are represented in Figure 6e. Again, there is a reciprocal relationship between
26 drug and light dose, with excellent phototoxicity in the $2.0\text{-}4.0 \mu\text{M}$ drug concentration
range.

The activity of HBEA-R1 and HBBA-R2 activity is effected by oxygen levels
(Figure 10). Both of these compounds are able to kill cells in the absence of oxygen or
at low oxygen levels. At the low oxygen levels higher doses of light are required to kill

1 cells than at higher oxygen levels. Since these compounds can kill cells at low oxygen
levels these compounds are suitable for the treatment of solid tumors, wherein hypoxia
may be present.

Example 15. Site-specific Modification of parent structures

6 The basic structures of the parent hypocrellins render them amenable to site -
specific modification [Diwu and Lown 1990]. A major advantage of hypocrellins as
photosensitizers for photodynamic therapy, rests in their ability to be synthesized in
pure, monomeric form. This feature significantly facilitates studies on phototoxic
mechanisms *in vitro*; however a major advantage is foreseen in the simplicity with
11 which pharmacokinetic studies may be executed. The low molecular weight facilitates
rapid distribution among tissues. The octanol-water partition coefficient is amenable to
site- specific alteration, a feature which will expedite synthesis of a series of derivatives
of graded hydrophobicity. For example, addition of the terminal hydroxyls and the
quaternary nitrogen atoms in the sidechains of HBEA-R1 and HBDP-R1, respectively,
16 may promote water solubility. This property will affect association with plasma
proteins and lipoproteins, and therefore, the tumor and normal tissue distribution and
kinetics of uptake and clearance. Butylamino- substitution of the phenolic hydroxyls,
yielding HBBA-R2 and HBBA-R1 has raised the extinction coefficient (ϵ) over six-fold
compared with HB, with a concomitant 7-10-fold enhanced phototoxicity. The same
21 substitution resulted in at least a 5-fold reduction of cytotoxicity, compared with the
parent compound. The ethanolamine substitution resulting in HBEA-R1 has enhanced
 ϵ to a similar degree, with attendant heightened phototoxicity. Finally, 2-
N,Ndimethylamino)propyl- amination of HB's phenolic hydroxyl (HBDP-R1) resulted
in a 4.8-fold boost in ϵ , with augmented phototoxic activity. While these three
26 modifications actually reduced singlet oxygen yield to varying degrees, enhanced
phototoxicity may relate to altered intracellular sensitizer distribution, with advantageous
targeting of the phototoxic species. Of more than 25 hypocrellin derivatives screened to
date, all have shown acceptable levels of cytotoxicity *in vitro*. At least seven of the
compounds photopotentiate by a factor of 10 (Table 1), with four possessing

1 exceptional photopotentiating ability. Direct molar comparison of the hypocrellin
concentrations required to effect a given degree of cytotoxicity or photopotentiation,
with those for P-II® is impractical, since the molecular weight of the photosensitizing
component(s) of P-II® is not clearly defined. Considering the low molecular weights
6 characteristic of the hypocrellin derivatives, and their predominance as monomeric
forms, the inventors did not deem it necessary to adhere to the tradition of longer drug
pre-incubation times characteristic of some porphyrins. Pre-incubation kinetics studies
confirm that varying the pre-incubation time for intervals up to 24 hours has no
significant effect on the drug concentration required to exert 50% cytotoxicity or
photopotentiation of cell kill. Recent fluorescence confocal microscopy studies indicate
11 that for most compounds examined, uptake is complete within two-hour incubation
period. Kinetics studies are essential to avoid premature rejection of potential
photosensitizing compounds.

These *in vitro* clonogenic screening assays provide a method to screen for
compounds with better physicochemical properties (and ideally, cytotoxicity and
16 photopotentiation) of promising hypocrellin derivatives. This approach has been useful
and efficient to estimate sensitizer concentration ranges required to affect cytotoxicity
and photopotentiation. This method can be used to select compounds suitable for
preclinical and clinical evaluations.

Other mechanistic studies concern the effect of photosensitizer and PDT on the
21 cell cycle. Preliminary findings for HBEA-R1 and HBBA-R2 gave no indication of
major cell cycle perturbations in the presence or absence of light treatment. This
finding is important, since treatment induced synchrony or selective toxicity could
result in residual PDT-resistant tumor cell populations.

The inventors have also found evidence that both of these sensitizers promote
26 substantial apoptotic cell death following PDT. Control and treatment of EMT6/Ed
monolayer cells were subjected to propidium iodide staining and zero-integrated field
electrophoresis (ZIFE). Both techniques provided data implicating apoptosis as a major
contributor to PDT toxicity (Figure 11).

1 Example 16. In Vivo Studies

The inventors have performed *in vivo* studies in rodents to assess putative systemic toxicity and pharmacokinetic properties of selected compounds. These compounds are lipophilic and have a tendency to aggregate in aqueous solutions. The preferred routes of administration of drug not in a liposome formulation is either via
6 injection into or very near the tumor or intravenously (i.v).. After i.v. injection of HB into Balb/c EMT6/Ed cells, the maximum drug concentration in the tumor was highest between the time of injection and 2 hours after injection (Table 2). This timing of cellular uptake agrees with the *in vitro* data (Figure 9a.). The level of HB in the skin initially dropped quickly immediately after administration of the drug and the majority
11 of the drug is cleared from the body within 24 hours (Table 2).

Preliminary analysis of HBEA-R1 and HBBA-R2 indicates that these two compounds retain significant potency *in vitro* at 688 nm. Graded doses of HBEA-R1 and HBBA-R2 have been administered to Balb/c mice, to a maximum of 31 and 39 mg/kg body weight, (50 μ M) respectively. Acute and chronic toxicity studies in rats
16 with HBEA-R1 and HBBA-R2 demonstrate no significant effects to graded doses including 7.5 mg/kg total body weight 11-12 μ M. No problems of acute or chronic toxicity (60 days) were observed following total doses of 50 μ M of these drugs. Primary endpoints included behavioral and physiological signs, such as consitutive grooming, hunching, altered gait, lethargy, aggression, and breathing rate and body mass compared
21 with control animals.

Example 17. Laser Light Wavelength and Dosage

Both the concentration of drug and the dosage of light are important for treatment of tumors. Balb/c mice with EMT6/Ed tumors with 50 μ mol/kg body
26 weight of HBEA-R1 received various light dosages. The mice that received 100 Joules of 630 nm light (duration approximately 10 minutes) experienced approximately 90% tumor cure, mice that received 50 Joules of 630 nm of light experienced only a 40% cure rate and the cure rates were significantly lower at the lower light dosages (Figures 7 and 8).

1 This invention provides a method for treating cancer which is enhanced in the
presence of light wavelengths between 400 and 850 nm (see Figure 3 and Table 1 or
optimal wavelengths for individual compounds). The absorption spectra for many
compounds are included in Figure 3 and the main absorption peak for each compounds
are included in Table 1. Many of these compounds have significant absorbance around
6 the 630 nm (600 to 700 nm range) (Table 1). The optimal wavelength is different for
each compound (Table 1). For HBEA-R1 and HBBA-R2 wavelengths between at least
630 and 688 nm are capable of killing cells. For deeper or larger tumors the longer
wavelengths are preferred. For superficial tumors, laser wavelengths with lower
wavelengths or wavelengths in the green spectrum would be more suitable to use
11 (Nseyo et al., 1993) since the light does not penetrate as far. The ability of these
compounds to be photopotentiate at higher wavelengths increases the size of tissue that
can be treated with PDT and increases the depth at which treatment can be provided
using PDT. Fibre optic probes can be utilized to direct the laser light. Light may also
be delivered to a selected area, using an appropriate light source and shielding.

16 A method for treating bladder is described by Nseyo and associates (1993) this
method can be applied using the compounds described in Table 1 or Figure 2 and drug
doses described and wavelengths described herein.

For applications of drug to a localized region or with identifiable target antigens
there are several methods that are suitable for delivery, the delivery system are
21 comprised of drug- liposome formulations, drug -monoclonal antibody delivery systems
such as monoclonal antibody -liposome, or applied to exposed surfaces using a
standard lipophilic skin cream. The drug can be applied topically or the route of
delivery of the drug or drug and delivery system could be intravenous,
intraperitoneally, intrathecally, intravesically, by intratumoral injection or by oral
26 ingestion.

Example 18. Composition Delivery

These compounds can be delivered to cells via liposome-mediated drug delivery,
including conventional and "Stealth" liposomes, and immunoliposomes designed to

1 target the specific organ or cancer type. For example, liposomes containing one of the
compounds described in Figure 2 with a monoclonal antibody (mAb) 48-127 (from the
laboratory of Y. Fradet) attached to the surface could be used to target bladder cancers
(de Harven et al. 1992). 48-127 is a mAb which recognizes a surface glycoprotein found
on both human bladder transitional cell carcinoma (TCC) and AY-27 rat TCC.)
6 (Fradet et al. 1992, 1986; Fradet and Cordon-Cardo, 1993; Rao et al. 1993). An example
of how to prepare a stealth immunoliposome is as follows: liposomes containing the
particular drug can be prepared from hydrogenated soy phosphatidylcholine and
cholesterol and 5% PEG-DSPE (polyethylene glycol-diestearoyl
phosphatidylethanolamine) using the protocol described by Allen and associates (1991).
11 An antibody can then be covalently attached to the PEG terminus using a thioester
bond as detailed by Allen (1994). Many other methods for preparing liposomes and
immunoliposomes are described in the literature and patents and would be suitable
carriers for these drugs. Liposomes and immunoliposomes can be made containing one
or more of the compounds described herein using published procedures. Those with
16 ordinary skill can readily ascertain or determine the appropriate solvents, other
appropriate reagents and conditions without resorting to undue experimentation.

Example 19. Method for Identifying and testing new PDT photosensitizers

The desired characteristics for a PDT sensitizer comprise at least one or more of
21 the following characteristics: good absorption of light in a wavelength that penetrates
tissue to the desired depth (Absorption in the 600 nm to 850 nm range penetrate the
skin many mm), compound sensitive to pH - inactive, lower activity or activity
destroyed at the pH characteristic of normal tissues, but active or higher activity at the
pH of the cells or organisms to be treated; compound cleared from the body quickly
26 and if a compound is intended to treat solid tumors it should have the ability to
function either in the presence and/or absence of oxygen to address the problem of
tumor cell hypoxia. The photosensitizer should have low dark cytotoxicity, and
excellent photopotential of cellular damage. The PDT toxic effect may be mediated
via necrotic, apoptotic cell death, or by stasis of the tumor vasculature or vascular bed.

- 1 Example 20. Synthesis of Amino Acid Derivatives of Hypocrellin B for Conjugation Purposes
- a. To a cooled solution (ice-bath) of 1 (7.5g, 100 mmol) in 25 ml of aqueous 4 M NaOH solution was added slowly, and alternately, benzyl chloroformate (17g, 100 mmol) and 25 ml of aqueous 4 M NaOH solution (each in five portions),
6 over a period of 30 minutes. The resulting mixture was acidified with concentrated HCl, the precipitated product was filtered and dried. Recrystallization from chloroform gave the pure product (2) in 72% yield (15.1 g).
- 11 b. A solution of 2 (6.3 g, 30 mmol) in dichloromethane (100 ml) is cooled to -5°C (ice-salt bath), and concentrated H₂SO₄ (0.3 ml) is added. The solution is saturated with isobutene, and then stirred at room temperature for 48 hours. Excess isobutene was removed at the water pump and the organic layer was washed with water (2x, 100 ml), 5% aqueous sodium bicarbonate solution (2x,
16 100ml), water (50 ml), and dried (Na₂SO₄). Removal of the solvent *in vacuo* produced 3 as an oil, in 85% yield (6.8 g).
- c. Hydrogenation of 3 (7.95 g, 30 mmol) in 80 ml of absolute ethanol in the presence of 2.0 g of 5% Pd-C at room temperature and pressure was
21 accomplished in 3 hours. After removal of the catalyst, the resulting solution was concentrated *in vacuo* to about 30 ml, and phosphorus acid (2.5 g, 30 mmol) in 60 ml ethyl acetate was added. The organic layer was removed and evaporated to give unreacted 3 (0.4 g, 5%). The aqueous layer was made alkaline with NaOH (1.4 g), and then extracted with ethyl acetate (3x, 50 ml), dried
26 (Na₂SO₄) and evaporated *in vacuo* to produce 4 in 75% yield (2.97 g).
- d. A mixture of hypocrellin B (40 mg, 0.08 mmol) and 4 (50 mg, 0.4 mmol) was refluxed in acetonitrile for 24 hours. Removal of the solvent *in vacuo* and purification by silica gel column chromatography using chloroform:methanol

(60:1, v/v) as eluent produced product 5 in 70% yield (35 mg). IR, NMR, and FAB-MS indicate that the product is a mono-adduct.

e. To a solution of 5 (50 mg, 0.8 mmol) in dichloromethane (30 ml) is added slowly, trifluoroacetic acid (16 mg, 0.01 ml, 0.11 mmol), and the mixture was stirred at room temperature for 1 hour. The mixture was then extracted with dichloromethane (2x, 30 ml), washed with water, dried (Na_2SO_4) and removal of the solvent *in vacuo* produces 6. Purification by silica gel column chromatography using chloroform:methanol (30:1, v/v) as eluent produced product 6 in 80% yield (46 mg).

f. A mixture of 6 (50 mg, 0.083 mmol) and thionyl bromide (17.3 mg, 0.083 mmol) in 15 ml of dichloromethane is stirred at room temperature for 1.5 hours, and then the organic layer is separated, dried (Na_2SO_4) and removal of the solvent *in vacuo* produces 7. Purification by silica gel column chromatography using chloroform:methanol (50:1, v/v) as eluent produced pure product 7 in 75% yield (41 mg).

Example 21. pH dependency

The effects of the pH of the extracellular milieu were tested with respect to phototoxicity to determine whether the low pH characteristic of some tumors would enhance the phototoxic effect via enhanced sensitizer uptake. EMT6/Ed cells growing in monolayer culture were subjected to PDT as outlined above, with the exception that the pH of the culture medium was adjusted incrementally in the range of 7.5 — 5.5 with exogenously supplied lactic acid (0.0 - 4.0 mM) during the 2-hour photosensitizer incubation. Survival relative to untreated controls was determined by clonogenic assay.

The effect of altering the extracellular pH on phototoxicity in EMT6/Ed cells in monolayer culture is summarized for HB between 0.0 and 5.0 μM in Figure 12. In the absence of HB, manipulation of extracellular pH with lactic acid has negligible effect on cloning efficiency. The pH effect is most pronounced at an extracellular pH of 6.5 for all HB concentrations, with the maximal effect at 3.0 μM of the sensitizer in the presence of light. Under these conditions, cytotoxicity is potentiated by a factor of 4 to

1 5 with respect to that at physiological pH.

Example 22. Genotoxicity

Preliminary studies were conducted on HBBA-R2 to estimate mutagenic potential. The *Salmonella typhimurium* histidine reversion system was used to detect
6 frame shift mutations (strain TA98) or base substitutions (strain TA100) in DNA, according to Organization for Economic Cooperation and Development (OECD) Guidelines 471 and 472. The plate incorporation assay procedure (Ames assay) was used, with HBBA-R2 introduced into the culture mixture to a maximum of 1.0 mg/plate in 100 μ l of dimethyl sulfoxide (DMSO). This sensitizer concentration
11 showed detectable cytotoxicity in both test strains.

Table 3 below summarizes results of experiments designed to identify and characterize putative genotoxic properties of the photosensitizer HBBA-R2. Data are indicative of properties of HBBA-R2 incubated with two strains of *Salmonella typhimurium*, TA98 and TA100, in the dark. These strains were chosen for their
16 propensity to detect a wide range of compounds with mutagenic potential. The negative control for both strains was DMSO (solvent) applied in the same concentration as that used for the test compounds. No mutagenic activity was observed for DMSO. Positive controls included 2-nitrofluorene, sodium azide, and 2-aminofluorene. They demonstrated the expected pattern of mutagenic activity in the presence and absence of
21 mammalian metabolic activation system (S9) in each strain. HBBA-R2 did not demonstrate any mutagenic activity in either strain, at concentrations to and including 1000 μ g/plate, indicating that at clinically relevant concentrations, HBBA-R2 promotes neither base substitution, nor frame -shift mutations.

The absence of genotoxicity in these mutagenicity studies may be related to the
26 finding that the free drugs are not found in the cell nucleus, within the detection limits of CLSM, or that they are not inherently genotoxic. Conjugates that enter the nucleus, e.g., PQPs conjugated to DNA minor groove binding agents, were not tested. Nuclear labeling may provide augmented phototoxicity through DNA targeting, with no added burden of cytotoxicity.

Table 1. Physical and Chemical Properties for Hypocrellins of Use in PDT

Name of Compound (structure no. in Figure 2)	Chemical Formula	F.W.	Abs. Peak in red Spectral region (nm)	A _{max} /solvent	A ₄₃₀	Extinction Coefficient (x 10 ³) (630 nm)	¹ O ₂ Yield	LD ₅₀ Dark (μM)	LD ₅₀ Light (μM)	Photo- potenti- ation Factor
HA (1)	Hypocrellin A									
	C ₃₀ H ₂₈ O ₁₀	546	658*	0.093/DMF	0.086	0.86	0.84	15	3-5	3-5
HB (5)	Hypocrellin B									
	C ₃₀ H ₂₈ O ₉	528	658*	0.118/DMF	0.100	1.00	0.74	20	1.5-2	10-13
HA-Mg ⁺⁺	HA-Mg ⁺⁺ (Ac) ₂									
	C ₃₄ H ₃₂ O ₁₂ Mg	652	616	0.958/EtOH	0.447	4.47	0.71	>25	>5	ND
HB-Mg ⁺⁺	HB-Mg ⁺⁺									
	C ₃₄ H ₃₂ O ₁₁ Mg	634	622	0.604/EtOH	0.527	5.27	0.53	10	1	10
DAHA (2)	Decetylated-HA									
	C ₃₄ H ₃₈ O ₁₀ Mg	592	622	0.651/EtOH	0.570	5.70	0.51	>25	>5	ND
HBAC-R1	Cystamine-HB									
	C ₃₃ H ₃₇ O ₈ Mg	585	646	0.417/CHCl ₃	0.388	3.88	0.40	12.5	1	12.5
HBAC-R2	Cystamine-HB									
	C ₃₃ H ₃₇ O ₈ Mg	585	600	0.337/DMSO	0.244	2.44	0.31	12.5	5	2.5
HBBA-R2 (6)	n-butylaminated HB									
	C ₃₄ H ₄₀ N ₂ O ₇	780	616*	0.628/CHCl ₃	0.619	6.19	0.32	>100	0.2-0.6	167-500
HBAM-R1	2-morpholino-ethyl- aminated-HB									
	C ₃₄ H ₄₀ N ₂ O ₇	752	658				0.70	>25	4	>6.25
HBDD-R2	2-(N,N-diethyl-amino) ethylamine-HB									
	C ₃₄ H ₄₂ N ₂ O ₇	696	646*	0.508/CHCl ₃	0.055	0.55	0.36	>25	7.5	>3.3
HBDDP-R1 (7)	2-(N,N-dimethyl- amino) propylamine- HB									
	C ₃₆ H ₄₄ N ₂ O ₇	724	640*	0.463/CHCl ₃	0.480	4.80	0.42	>25	0.5-1.5	>16.6-50

Name of Compound (structure no. in Figure 2)		Chemical Formula	F.W.	Abs. Peak in red Spectral region (nm)	A _{max} /solvent	A ₃₀	Extinction Coefficient (x 10 ⁴) (630 nm)	¹ O ₂ Yield	LD ₅₀ Dark (μM)	LD ₅₀ Light (μM)	Photo- potenti- ation Factor
HBEA-R1 (3)	Ethanolamine-HB	C ₂₄ H ₃₄ N ₂ O ₆	614	696*	0.625/DMSO	0.623	6.23	0.60	15-25	0.15	100-167
HBEA-R2 (8)	Ethanolamine-HB	C ₂₄ H ₃₄ N ₂ O ₆	614	634*	0.162/DMSO	0.127	1.27	0.70	25	7.5	3.3
HBED-R2	Ethylenediamine-HB	C ₂₄ H ₃₂ N ₄ O ₆	696	614*	1.449/DMSO	1.239	12.39	0.50	>25	3-5	5-8.3
HBM-A-IV	Methylamine-HB	C ₂₀ H ₃₀ N ₂ O ₆	696	640	0.246/CHCl ₃	0.246	2.46	0.80	8.5	1	8.5
DBHB	5,8-dibromo-HB	C ₂₂ H ₂₂ O ₆ Br ₂	531	ND	ND	ND	ND	0.62	10	3	3.3
DMF-HB	demethylated HB	C ₂₂ H ₂₄ O ₆	686	648*	0.469/EtOH	4.77	4.77	0.42	>25	3-5	>5-8.3
JL-1-1 (4)		C ₂₀ H ₂₄ O ₁₂	578	594	0.478/CHCl ₃	0.062	0.62	0.72	>70	2-4	>18.5

HBBA-R2, HBDA-R1, HBEA-R1, and JL-1-1 demonstrate average or lower than average toxicity, with excellent potentiation. For the purposes of this study, the LD₅₀ light dose was not fixed. For the compounds tested, this dose is 0.75 - 1.0 J/cm² of 630 nm light.

ND = not done. * Significant light absorption at 630 nm. F.W. = formula weight.

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Table 2.Tissue Uptake of ^{14}C -Hypocrelin B (dpm/g)

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Tissue	0 Hours	2 Hours	24 Hours	48 Hours
Heart	113,920 \pm 3,365	5,135 \pm 910	7,835 \pm 1,810	2,325 \pm 245
Lung	651,100 \pm 42,668	8,580 \pm 655	3870 \pm 525	2,975 \pm 360
Fat	20,550 \pm 715	38,570 \pm 5,610	19,550 \pm 2,210	19,335 \pm 2,335
Liver	394,190 \pm 7,540	24,620 \pm 4,885	22,495 \pm 4,440	9,215 \pm 720
Spleen	151,870 \pm 9,395	58,900 \pm 4,205	14,970 \pm 3,215	26,700 \pm 11,105
Stomach	28,280 \pm 145	21,630 \pm 3,345	34,385 \pm 8,795	12,460 \pm 975
Pancreas	32,010 \pm 2,165	13,185 \pm 12,055	32,390 \pm 11,840	16,915 \pm 3,845
Ileum	45,400 \pm 3,600	20,280 \pm 2,850	5,800 \pm 645	2,840 \pm 595
Kidney	67,344 \pm 950	20,855 \pm 3,955	12,050 \pm 1,845	4,535 \pm 765
Skin	14,970 \pm 74	3,130 \pm 221	2,700 \pm 170	1,590 \pm 250
Bone	19,825 \pm 2,300	3,955 \pm 2,070	660 \pm 215	1,125 \pm 310
Brain	17,560 \pm 560	3,855 \pm 170	2,840 \pm 275	845 \pm 90
Muscle	13,665 \pm 600	4,050 \pm 940	2,875 \pm 560	1,015 \pm 205
Tumor	7,885 \pm 270	3,775 \pm 400	2,950 \pm 80	2,165 \pm 470
Serum	69,975 \pm 1,925	1,655 \pm 170	1,020 \pm 160	700 \pm 240

21

26

Table 3

Mutagenicity of HBBA — R2 in Strains TA98 and TA100 of *Salmonella typhimurium*

Control/Test Compound	Concentration of Compounds, $\mu\text{g}/\text{plate}$	Number of Histidine Revertants per Plate			
		TA98		TA100	
Negative control: 100 μl DMSO/ plate	0	-S9	+S9	-S9	+S9
		18 \pm 1	27 \pm 2	133 \pm 4	157 \pm 5
HBBA — R2	8	47 \pm 13	41 \pm 1	119 \pm 9	117 \pm 13
	40	29 \pm 5	52 \pm 4	124 \pm 6	140 \pm 13
	200	25 \pm 2	29 \pm 1	160 \pm 3	148 \pm 10
	1000	16 \pm 2	13 \pm 2	125 \pm 16	132 \pm 12
Positive Controls					
2-nitrofluorene	5.0	1197 \pm 53	-	-	-
Sodium azide	1.5	-	-	2017 \pm 36	-
2-aminofluorene	10.0	-	2951 \pm 390	-	2601 \pm 321
\pm S9: Presence or absence of mammalian metabolic activation system					

While the invention has been described in some detail by way of illustration and example, it should be understood that the invention is susceptible to various modifications and alternative forms, and is not restricted to the specific embodiments set forth. It should be understood that these specific embodiments are not intended to limit the invention but, on the contrary, the intention is to cover all modifications, equivalents, and alternatives falling within the spirit and scope of the invention.

Claims

What is claimed is:

1. A method of treatment comprising administering a conjugate comprising a perylenequinone bound to a binding agent, and activating the perylenequinonoid by exposing the perylenequinone to light of a predetermined wavelength.

2. The method of claim 1 wherein the perylenequinone is selected from the group comprising hypocrellins, cercosporins, phleichromes, elsinochromes, cladochromes, erythroaphins, and calphostins.

3. The method of claim 2 wherein the perylenequinone is functionalized.

4. The method of claim 1 wherein the perylenequinone is a hypocrellin.

5. The method of claim 1 wherein the perylenequinone is non-toxic at high concentrations in its non-activated state and toxic at low concentrations in its activated state.

6. The method of claim 4 wherein the hypocrellin is a derivative of hypocrellin A or hypocrellin B.

7. The method of claim 6 wherein the hypocrellin B is functionalized.

8. The method of claim 7 wherein the functionalized hypocrellin B is conjugated to a binding agent.

9. The method of claim 8 wherein the hypocrellin B is non-toxic at high concentrations in its non-activated state and toxic at low concentrations in its activated state.

10. The method of claim 1 wherein the binding agent is selected from the group consisting of an antibody, a monoclonal antibody, or a fragment thereof.

11. The method of claim 10 wherein the binding agent is an antibody that binds to an antigen of a cancer cell.

12. The method of claim 11 wherein the cancer cell is selected from the group consisting of ovarian cancer, breast cancer, and gastrointestinal cancer.

1 13. The method of claim 1 wherein administering a conjugate comprises
administering a conjugate comprising a hypocrellin B derivative bound to a monoclonal
antibody or a portion thereof.

 14. The method of claim 1 wherein the binding agent binds to a DNA minor
groove.

6 15. A method of treatment comprising administering a conjugate comprising a
hypocrellin B derivative bound to a monoclonal antibody or antibody fragment, and
activating the hypocrellin B derivative by exposing the hypocrellin B derivative to light
of a predetermined wavelength.

 16. The method of claim 15 wherein the monoclonal antibody or antibody
11 fragment binds an epitope on a cancer cell, said cancer cell is selected from the group
consisting of ovarian cancer, breast cancer, and gastrointestinal cancer.

 17. The method of claim 15 wherein the hypocrellin derivative is non-toxic at
high concentrations in its non-activated state and toxic at low concentrations in its
activated state.

16 18. A composition comprising a perylenequinone conjugated to a binding agent.

 19. The composition of claim 18 wherein the perylenequinone is selected from
the group comprising hypocrellins, cercosporin, phleischromes, elsinochromes,
cladochromes, erythroaphins, and calphostins.

 20. The composition of claim 18 wherein the perylenequinone is a derivative of
21 hypocrellin A or hypocrellin B.

 21. The composition of claim 19 wherein the perylenequinone is non-toxic at
high concentrations in its non-activated state and toxic at low concentrations in its
activated state.

 22. The composition of claim 20 wherein the hypocrellin derivative is non-toxic
26 at high concentrations in its non-activated state and toxic at low concentrations in its
activated state.

1 23. The composition of claim 18 wherein the binding agent is a targeting agent
for a disease, disorder, malady, or condition.

 24. The composition of claim 18 wherein the binding agent is selected from the
group consisting of an antibody, a monoclonal antibody, or a fragment thereof.

 25. The composition of claim 24 wherein the binding agent is an antibody that
6 binds to an epitope of a cancer cell, said cancer cell is selected from the group consisting
of ovarian cancer, breast cancer, and gastrointestinal cancer.

 26. The composition of claim 18 wherein the binding agent binds to a DNA
minor groove.

 27. A composition comprising a hypocrellin derivative conjugated to a binding
11 agent.

 28. The composition of claim 27 wherein the hypocrellin derivative is non-toxic
at high concentrations in its non-activated state and toxic at low concentrations in its
activated state.

 29. The composition of claim 27 wherein the binding agent is a targeting agent
16 for a disease, disorder, malady, or condition.

 30. The composition of claim 27 wherein the binding agent is selected from the
group consisting of an antibody, a monoclonal antibody, or a fragment thereof.

 31. The composition of claim 27 wherein the binding agent is an antibody that
binds to an antigen of a cancer cell, said cancer cell is selected from the group consisting
21 of ovarian cancer, breast cancer, and gastrointestinal cancer.

 32. The composition of claim 27 wherein the binding agent binds to a DNA
minor groove.

 33. The composition of claim 27 wherein the hypocrellin derivative is a
derivative of hypocrellin B.

 34. A composition comprising a derivative of hypocrellin B conjugated to a
26 binding agent, said binding agent selected from the group consisting of an antibody, a

1 monoclonal antibody, or a fragment thereof; said binding agent binds to an antigen of a
cancer cell, said cancer cell is selected from the group consisting of ovarian cancer,
breast cancer, and gastrointestinal cancer; and wherein said derivative of hypocrellin B
is a functionalized derivative that can be activated by exposing the derivative to a pre-
determined wavelength of light.

6 35. The method of claim 1 wherein the method of treatment comprises treating
skin conditions, cancer, viral diseases, retroviral diseases, bacterial diseases, and fungal
diseases.

36. The method of claim 1 wherein the predetermined wavelength is between
about 400 nm and about 850 nm.

11 37. The method of claim 36 wherein the predetermined wavelength is between
about 600 nm and about 700 nm.

38. A method for destroying or inactivating tumor cells comprising
administering a suitable amount of at least one hypocrellin derivative, and activating the
hypocrellin derivative.

16 39. The method of claim 38 wherein activating the hypocrellin derivative
comprises exposing the hypocrellin derivative to a predetermined wavelength of light.

40. A method for treating a disease or condition comprising administering a
composition comprising a hypocrellin derivative and at least one of a pKa modifier, a
buffer, a salt, a base, an acid, saline, and an adjuvant.

21

Figure 1:

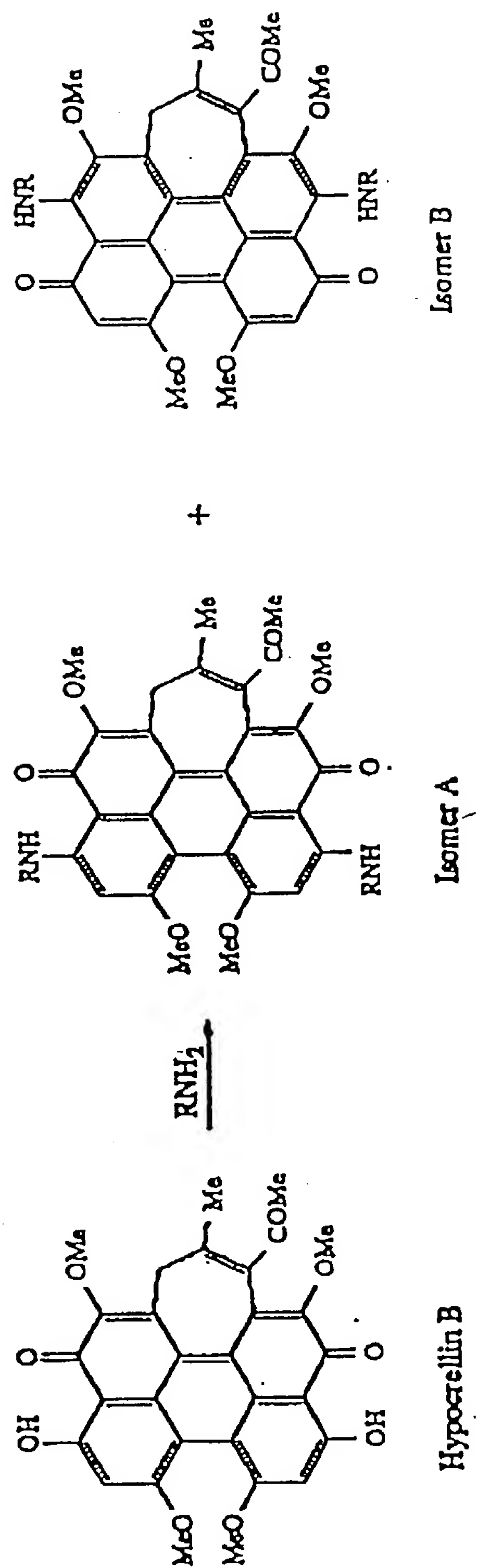
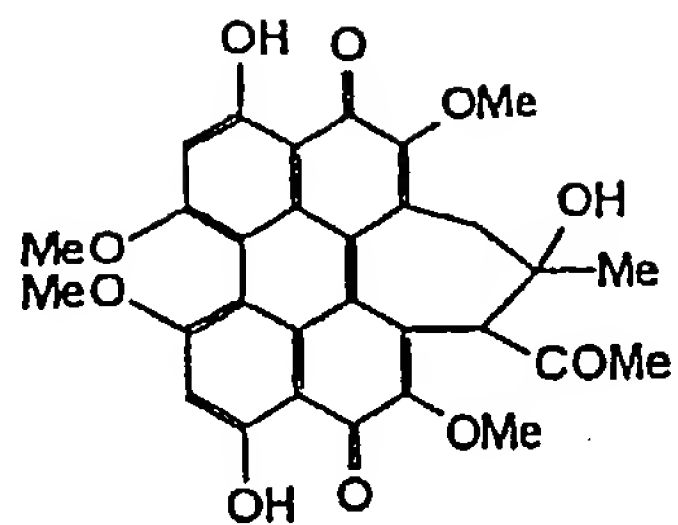
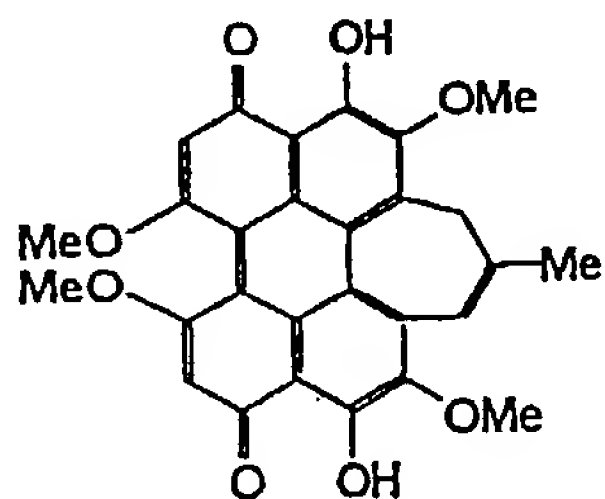


Figure 2: Chemical Structures

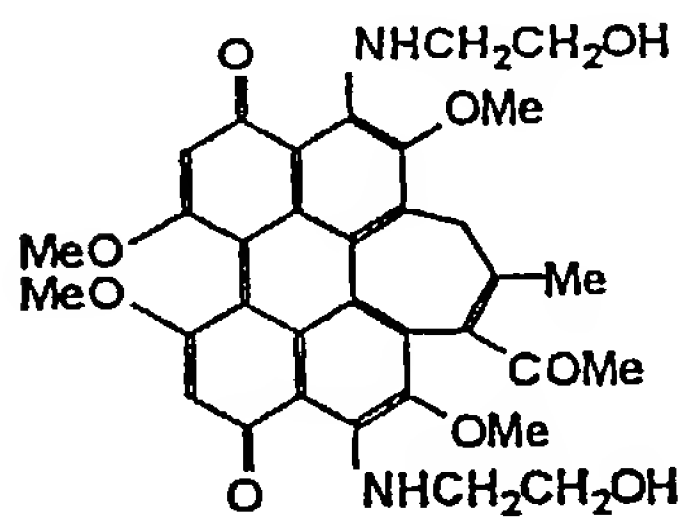
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HYPOCRELLIN A (HA)



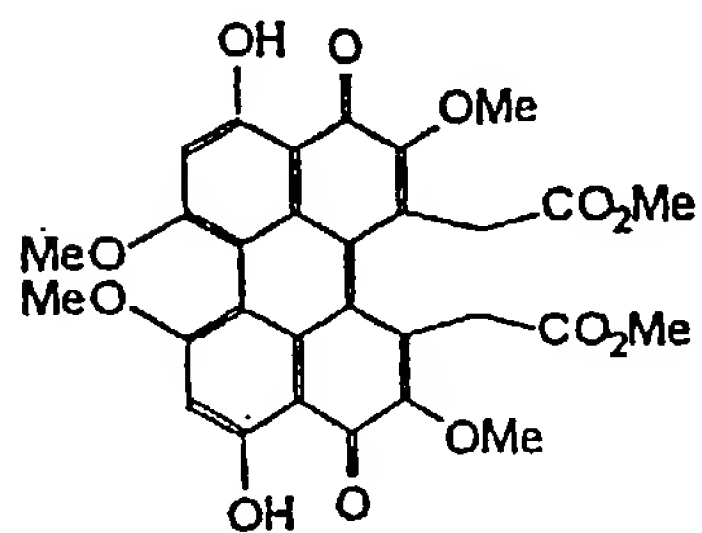
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DAHA



3

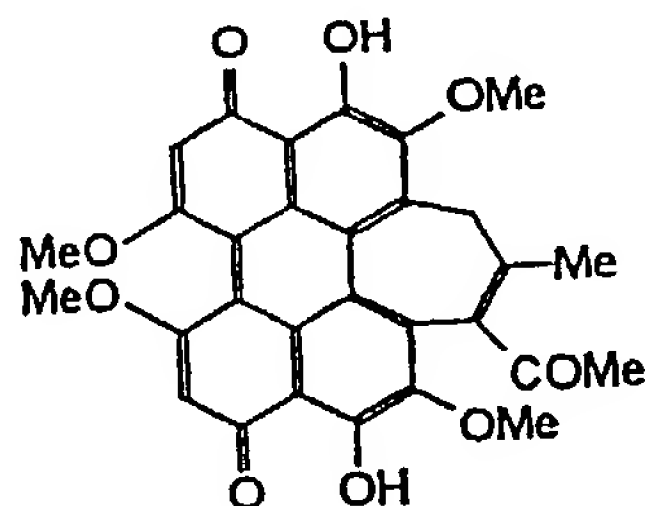
HBEA-R1



4

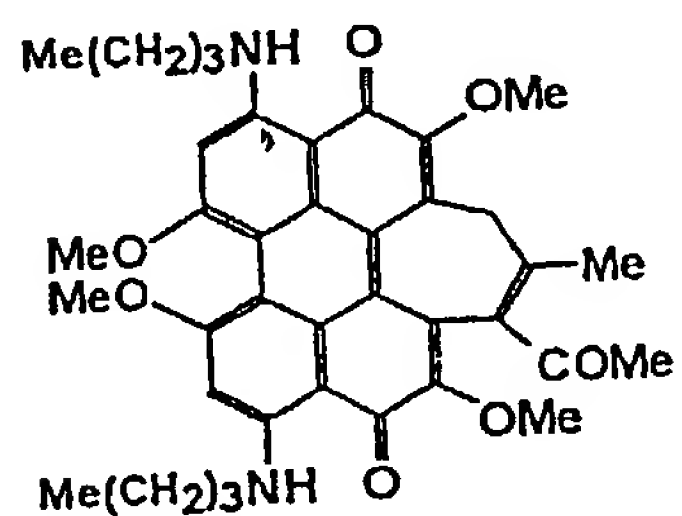
JL-1-1

Figure 2: (continued): Chemical Structures



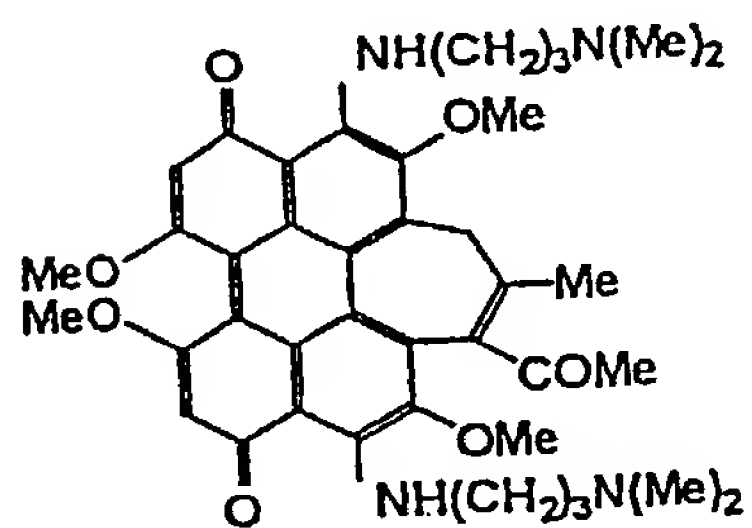
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HYPOCRELLIN B (HB)



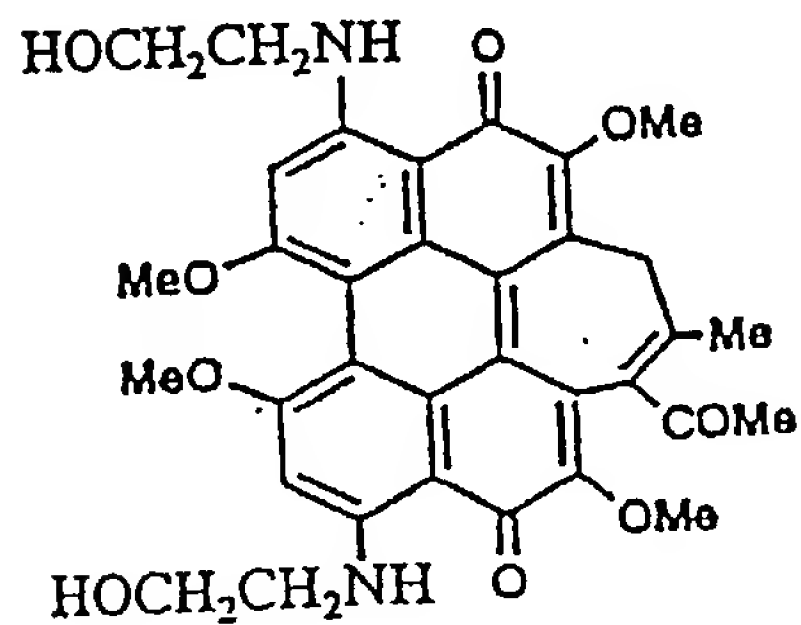
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HBBA-R2



7

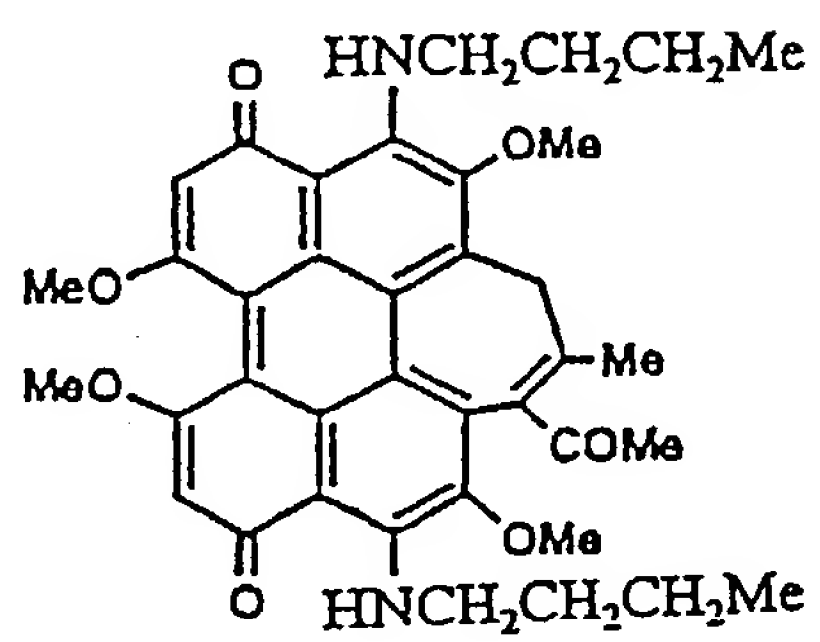
HBDP-R1



8

Compound 3A

Figure 2 (continued): Chemical Structures



Compound 4B

Figure 3:

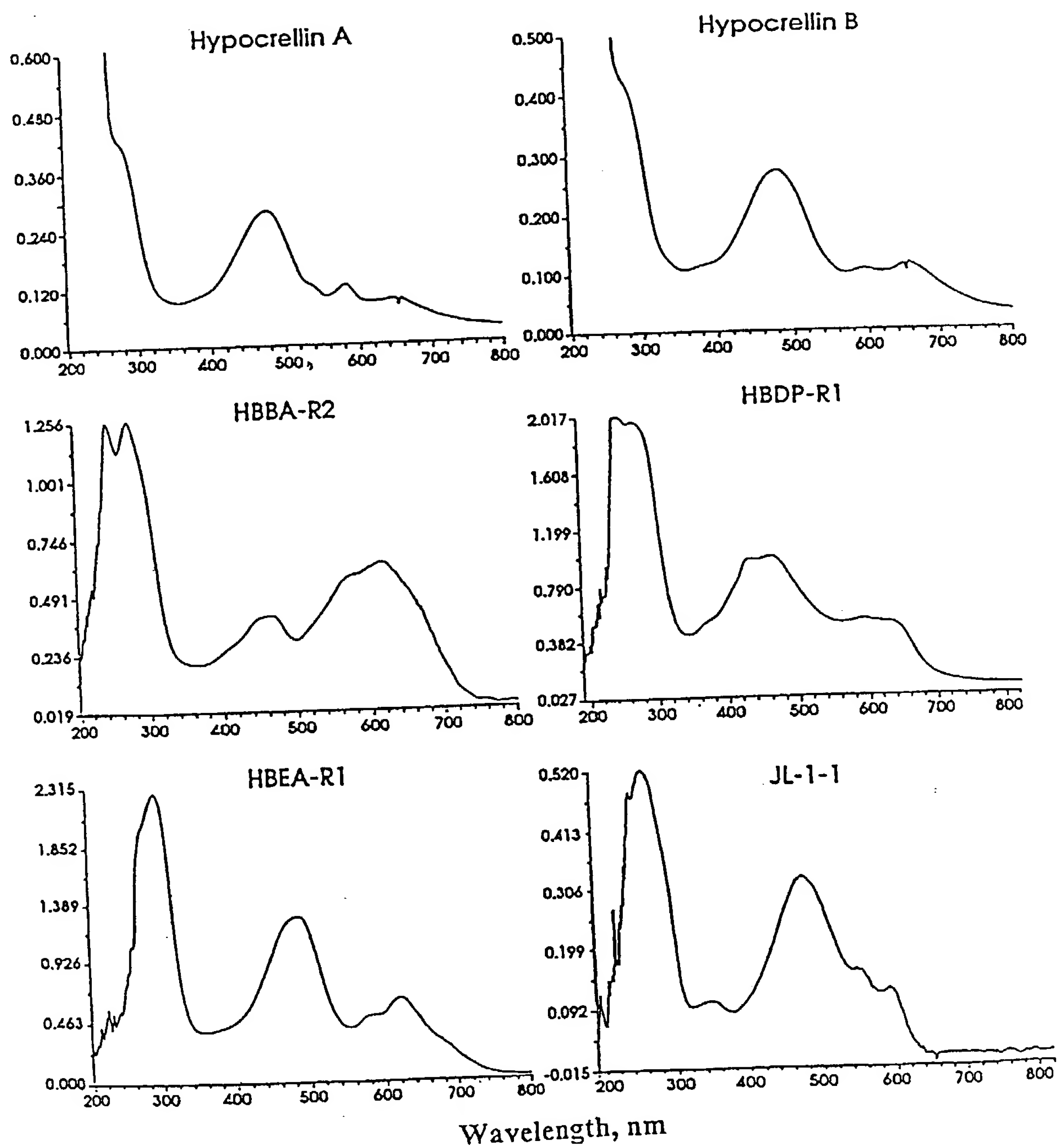


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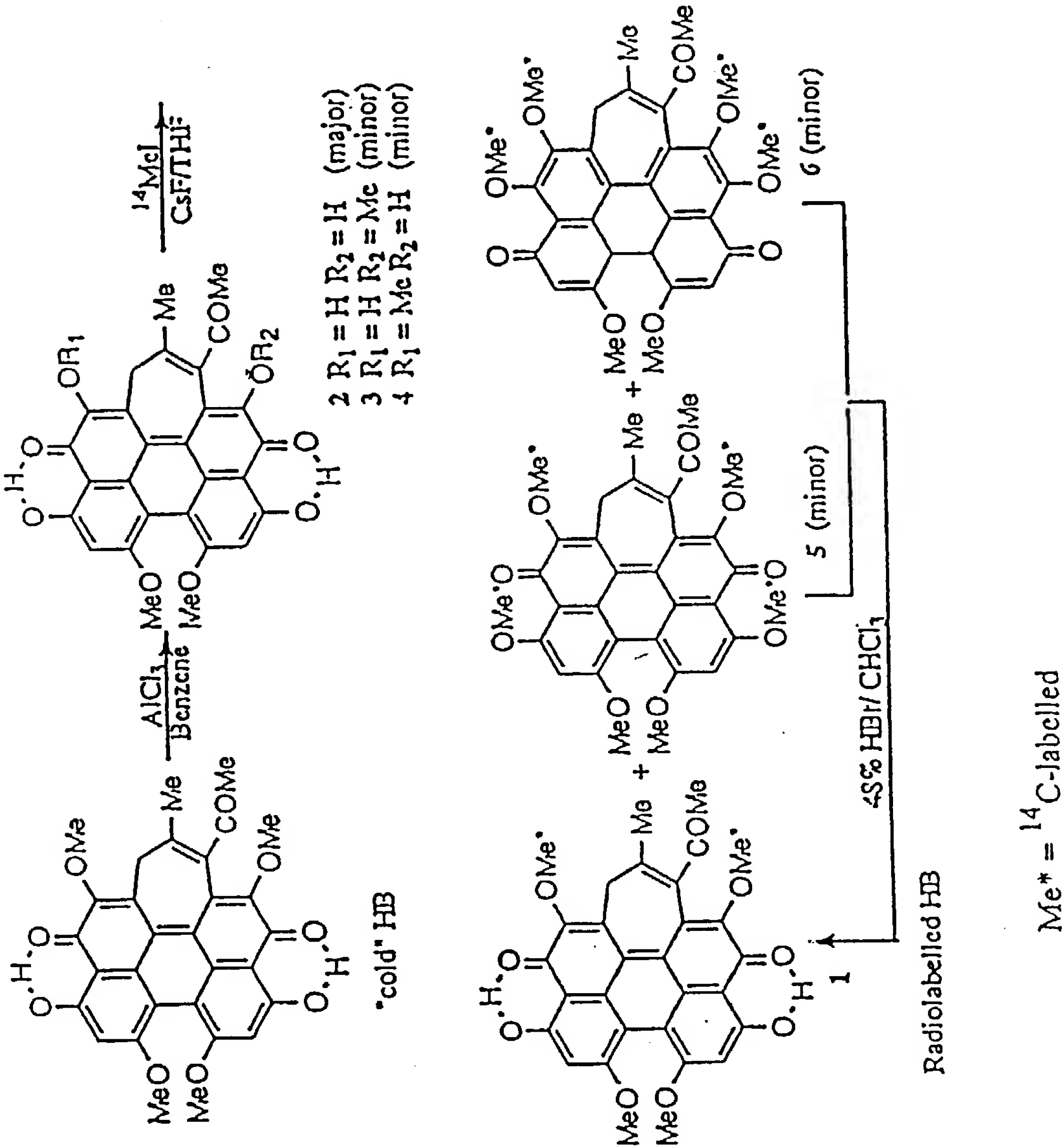


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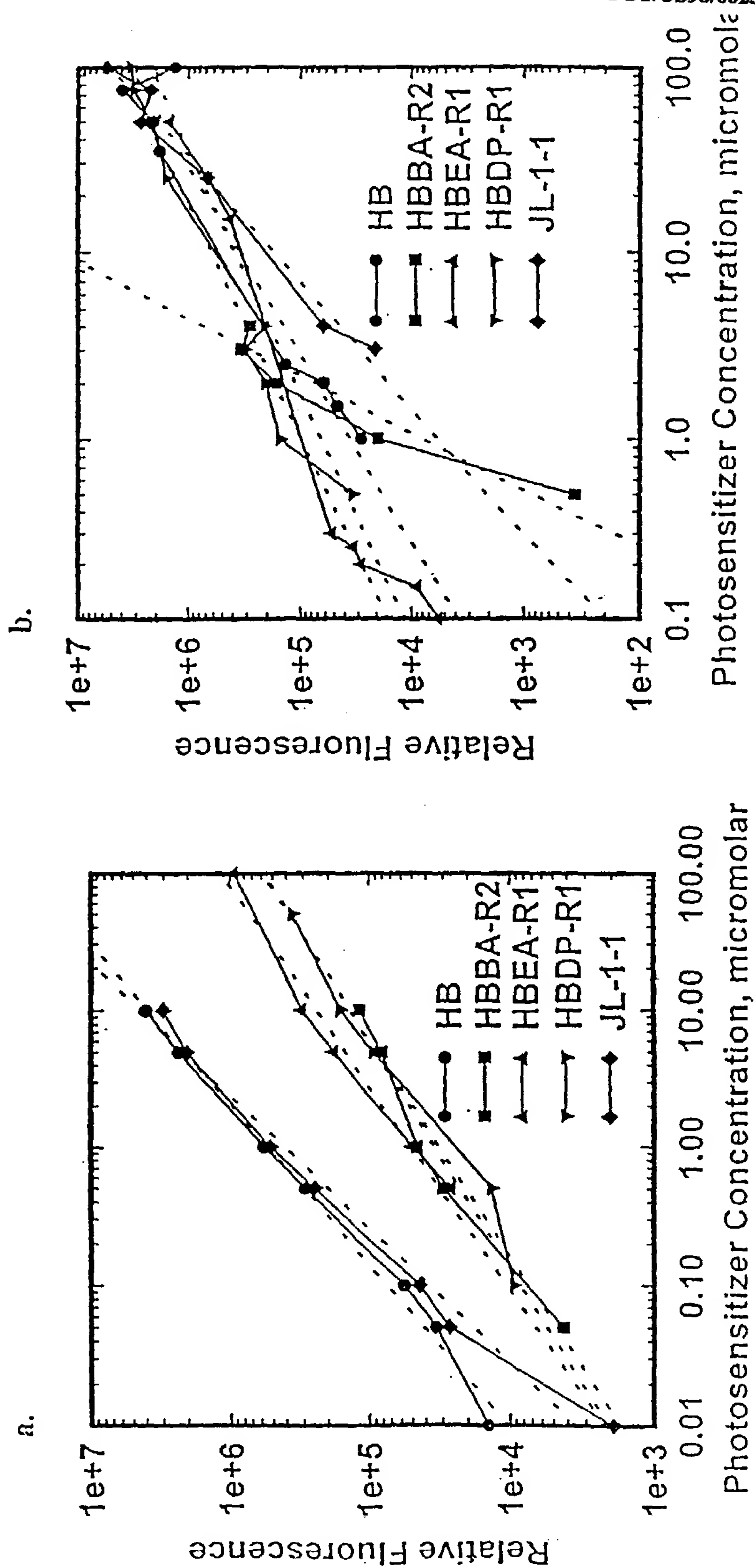
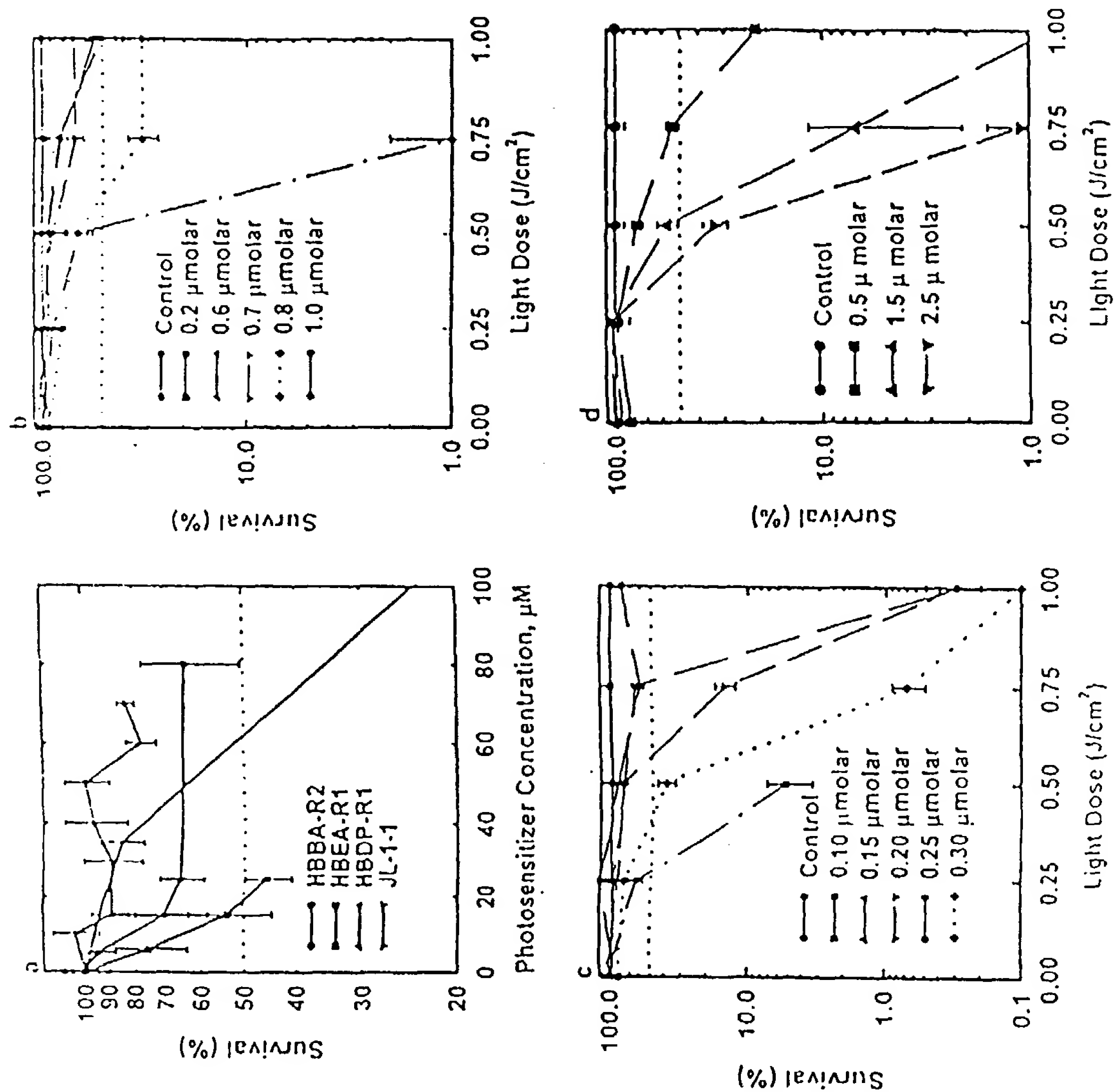


FIGURE 6



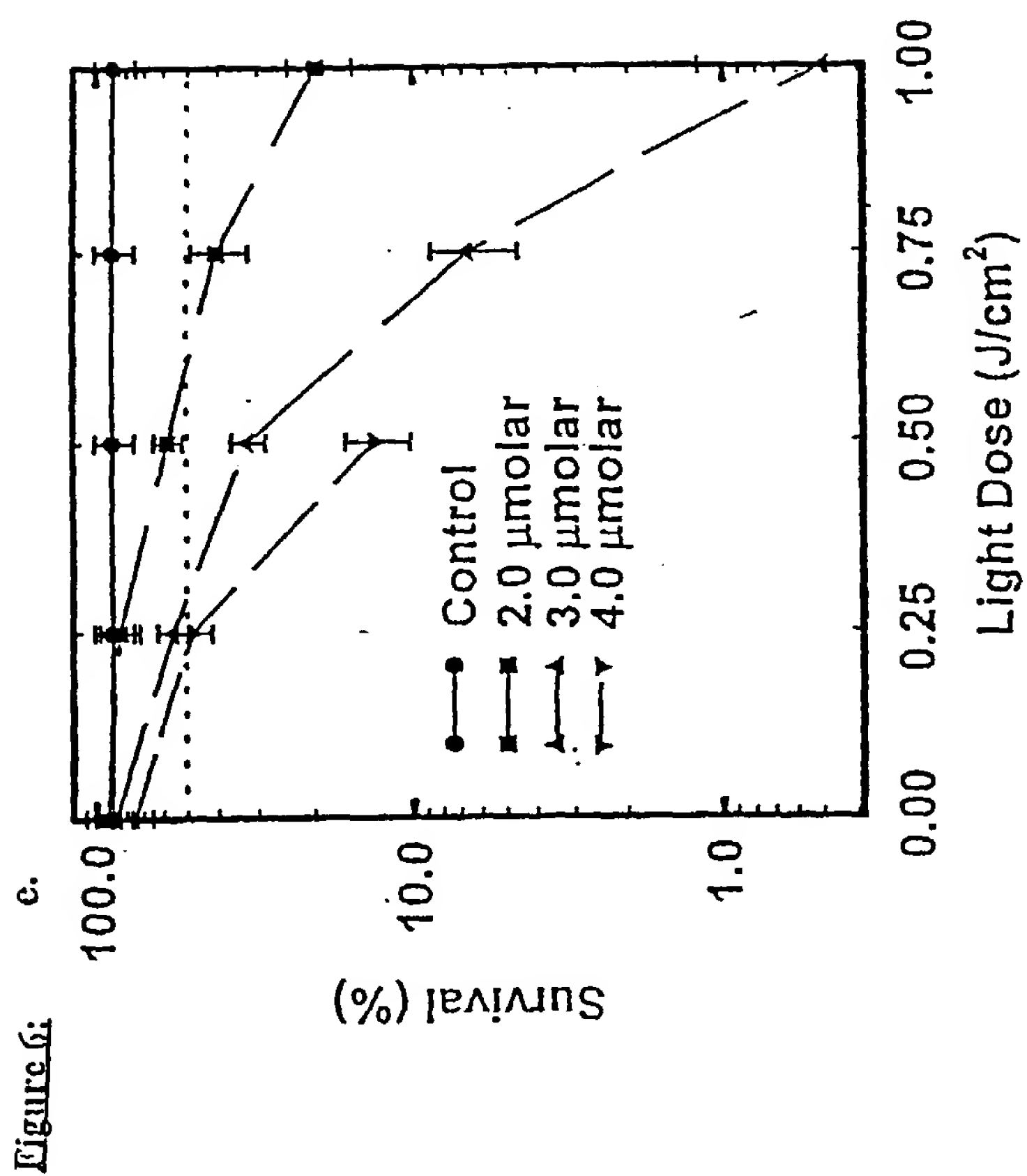


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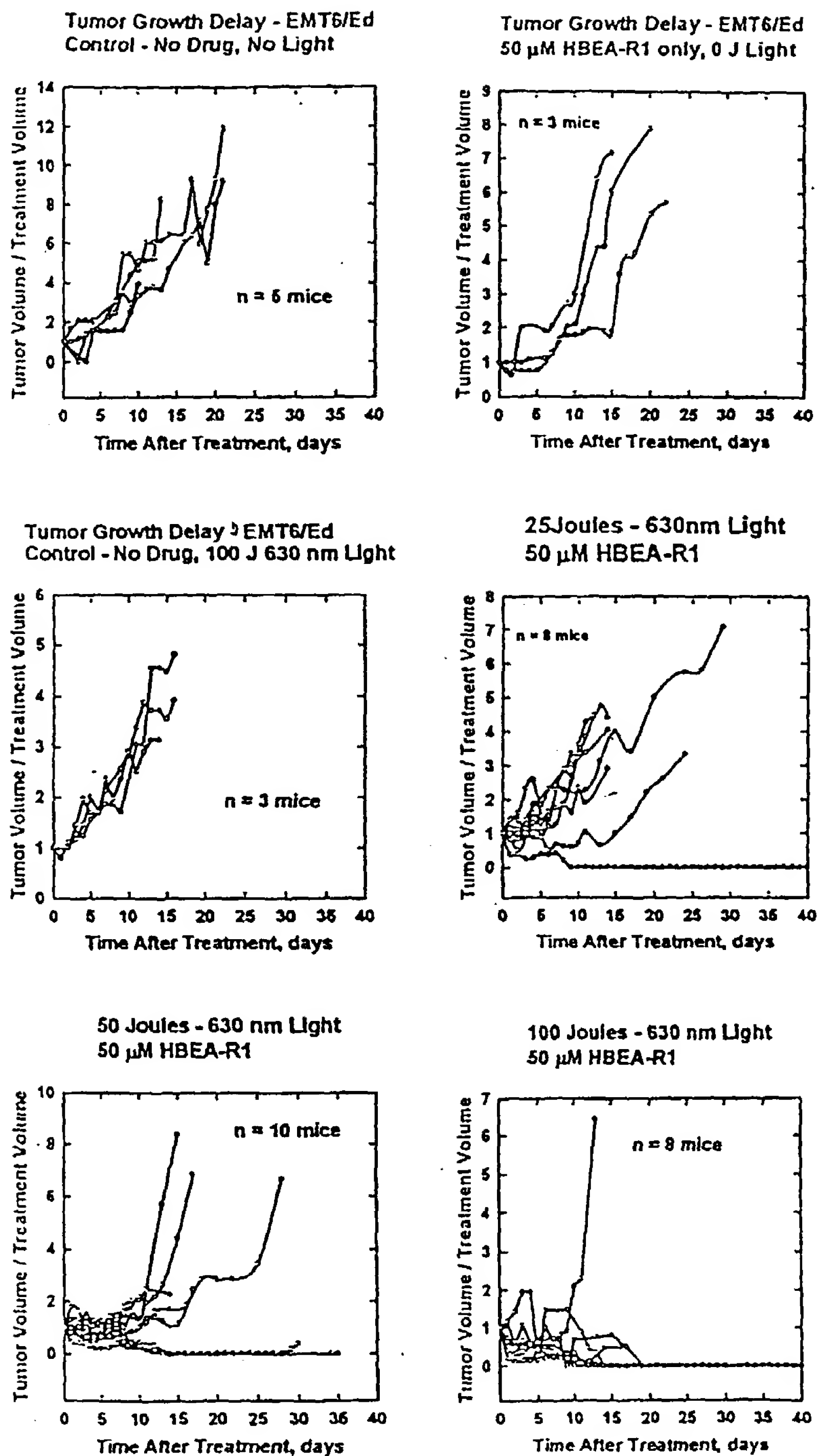
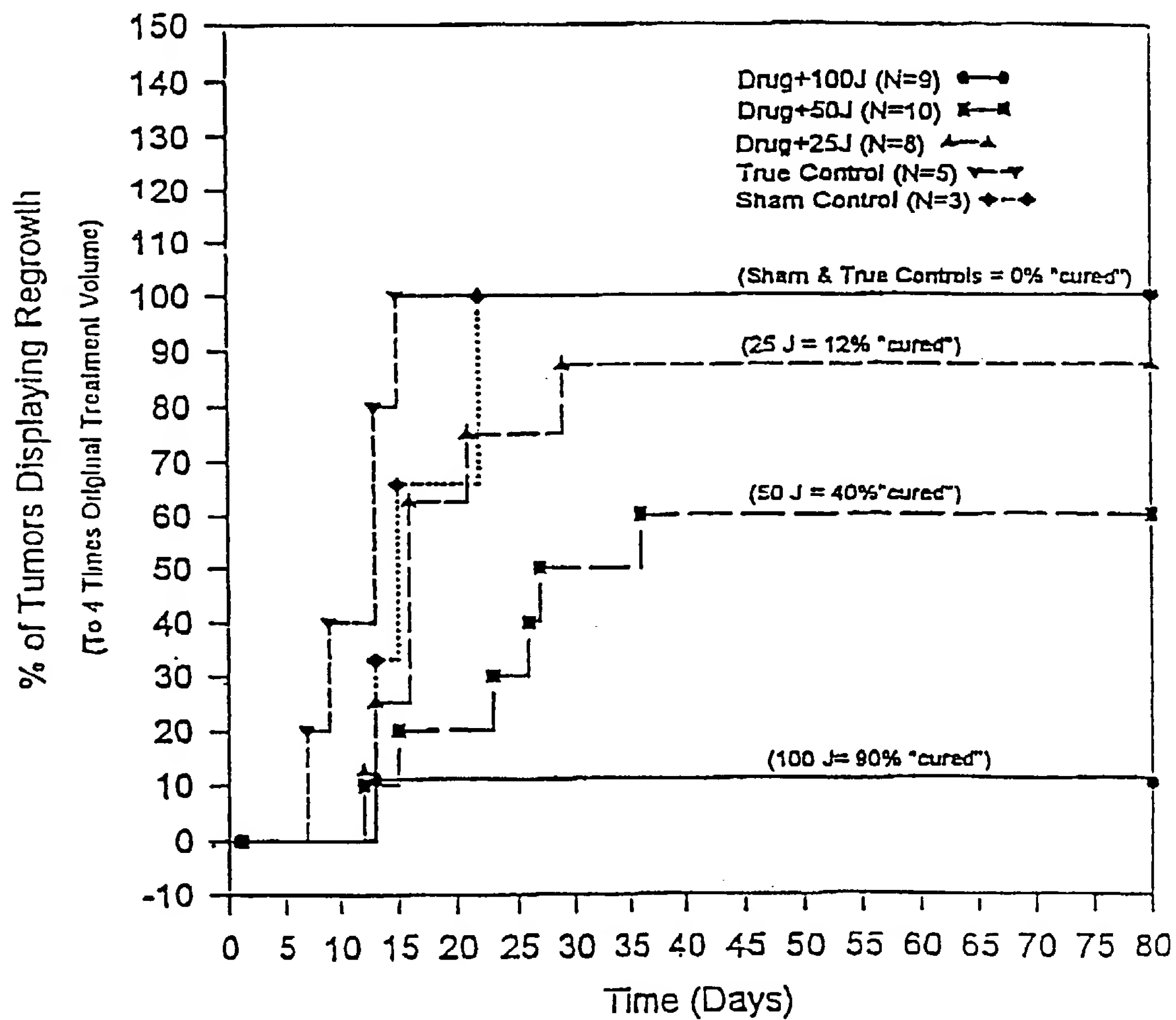
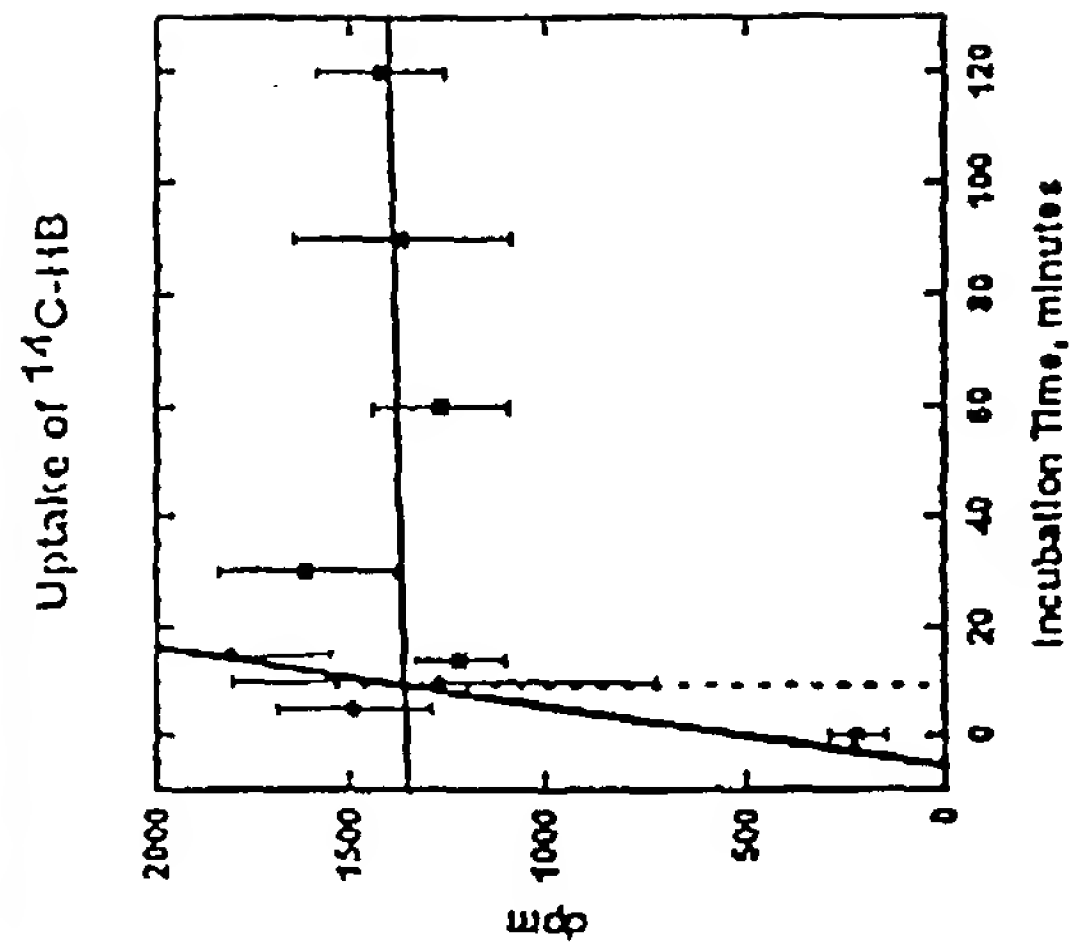


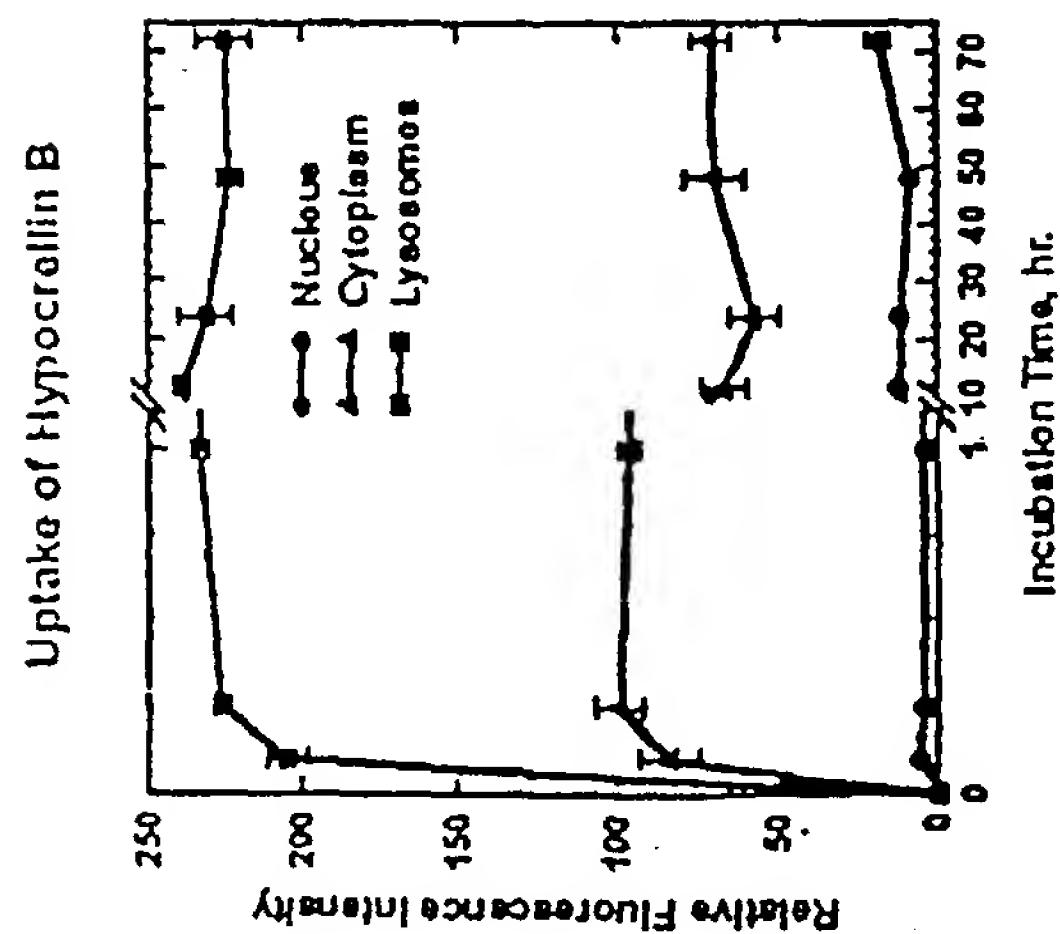
Figure 8:

Tumor Control After HBEA-R1 Administration

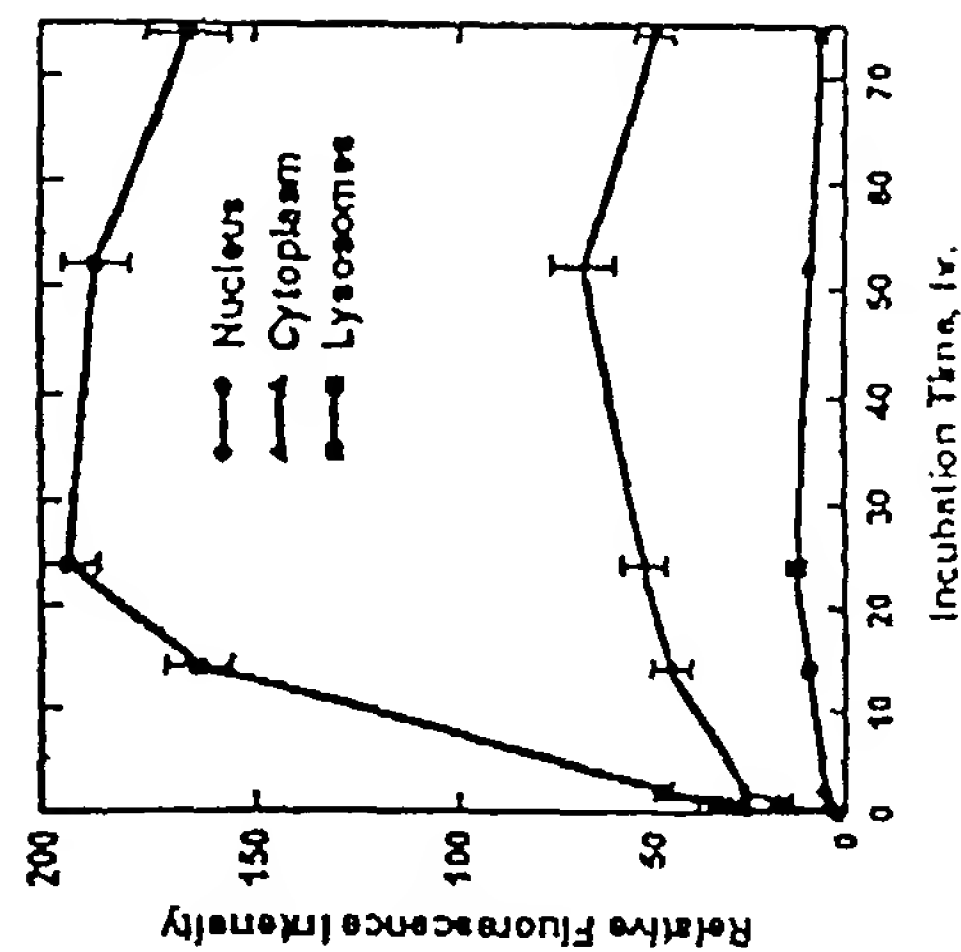




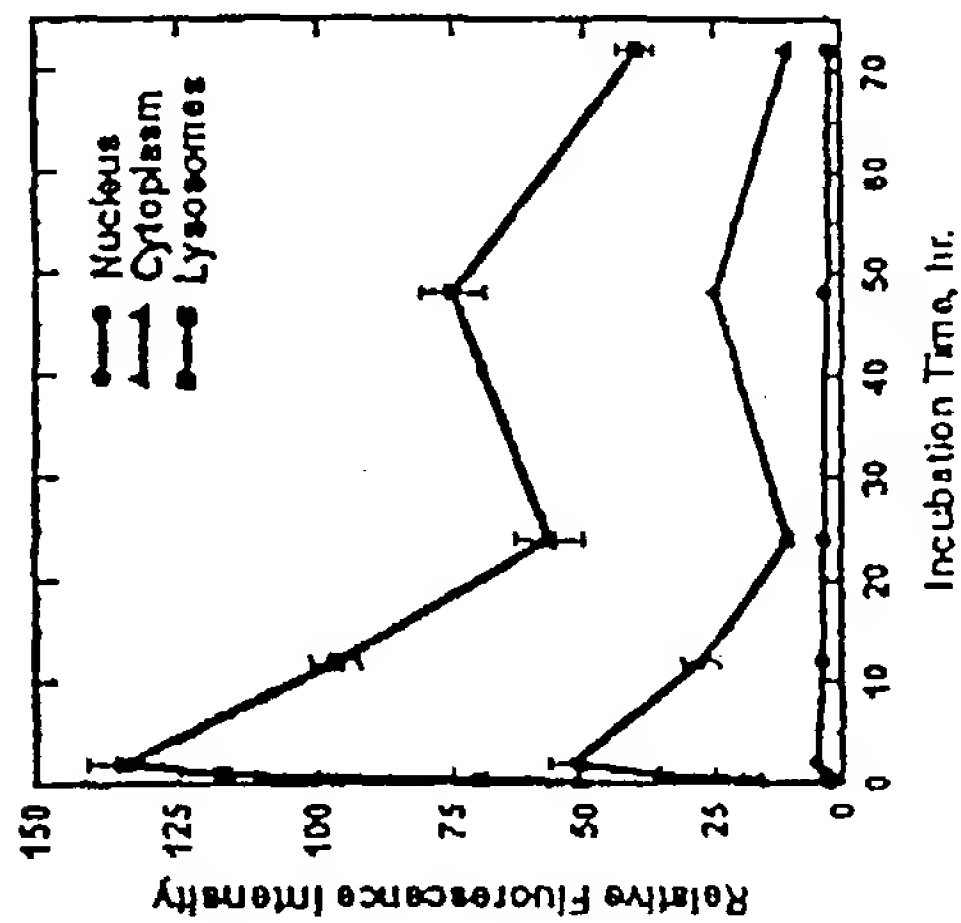
b.



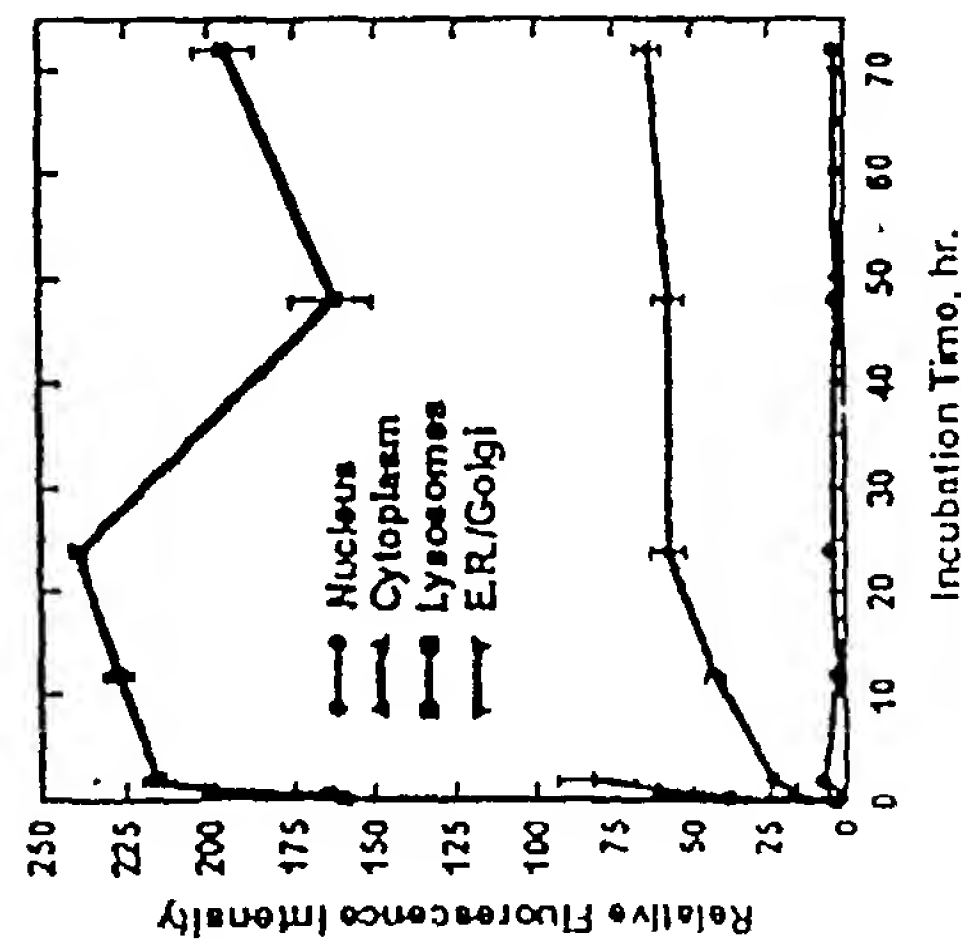
a.



c.



d.

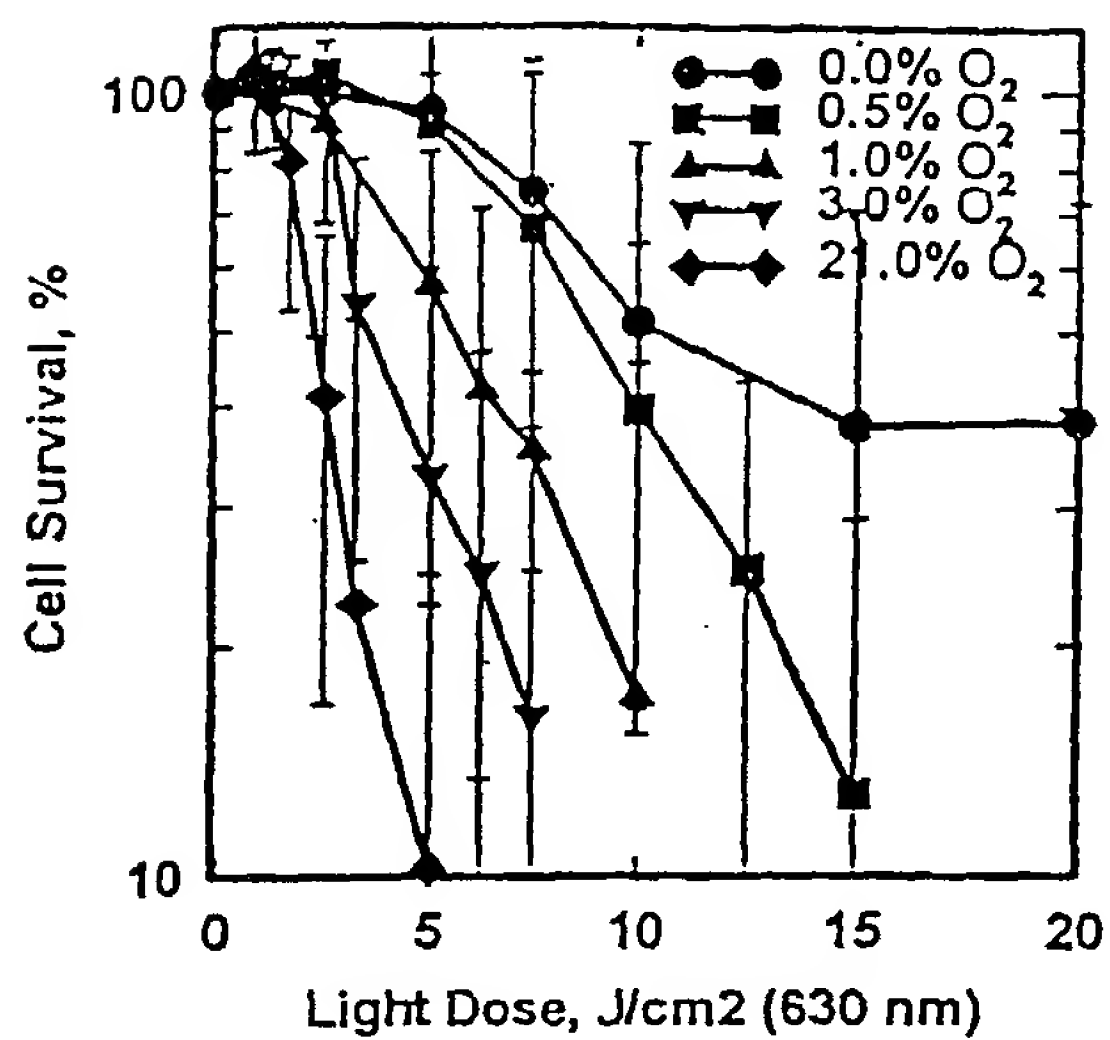


c.

Figure 9:

Figure 10:

Oxygen Dependency of HBEA-R1



Oxygen Dependency of HBBA-R2

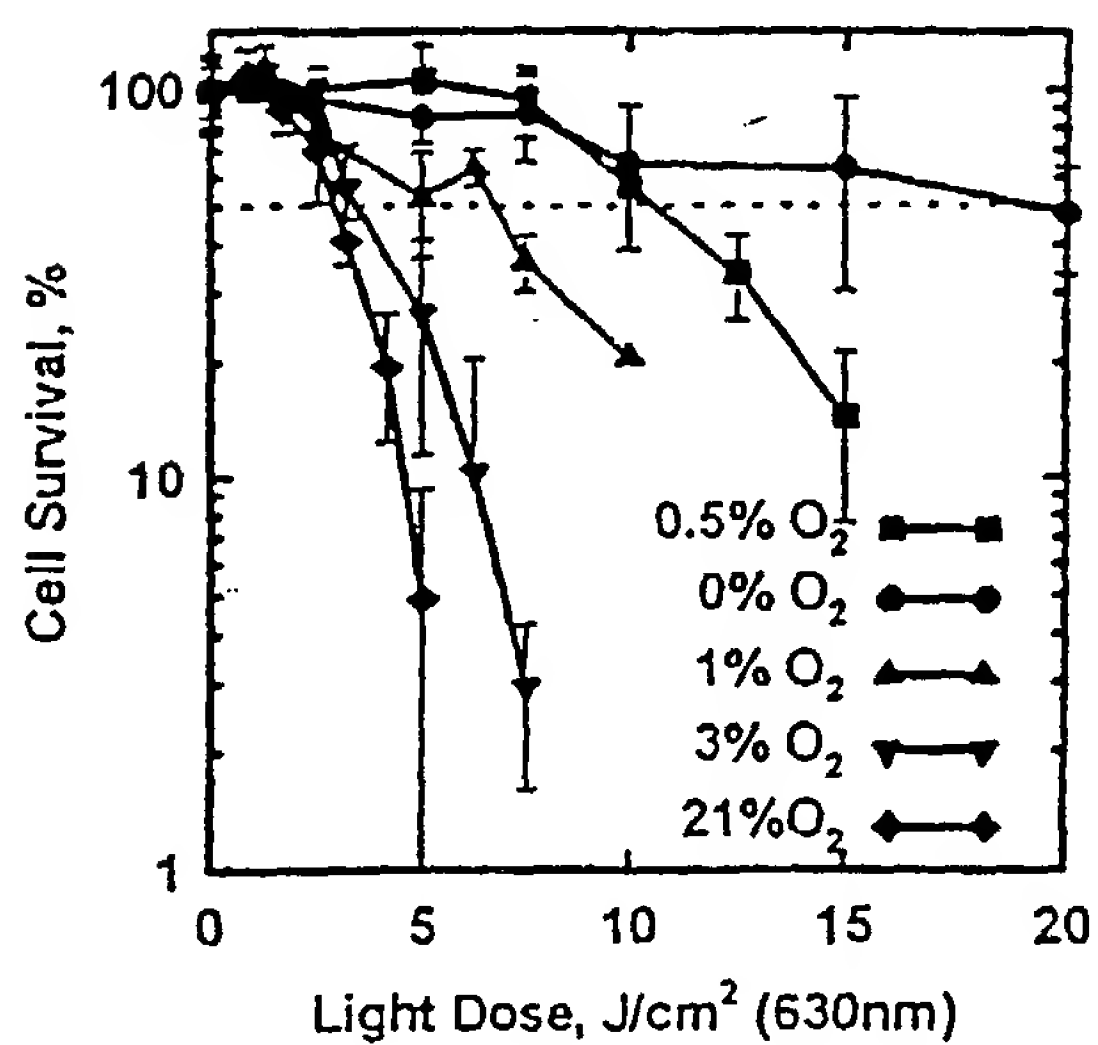
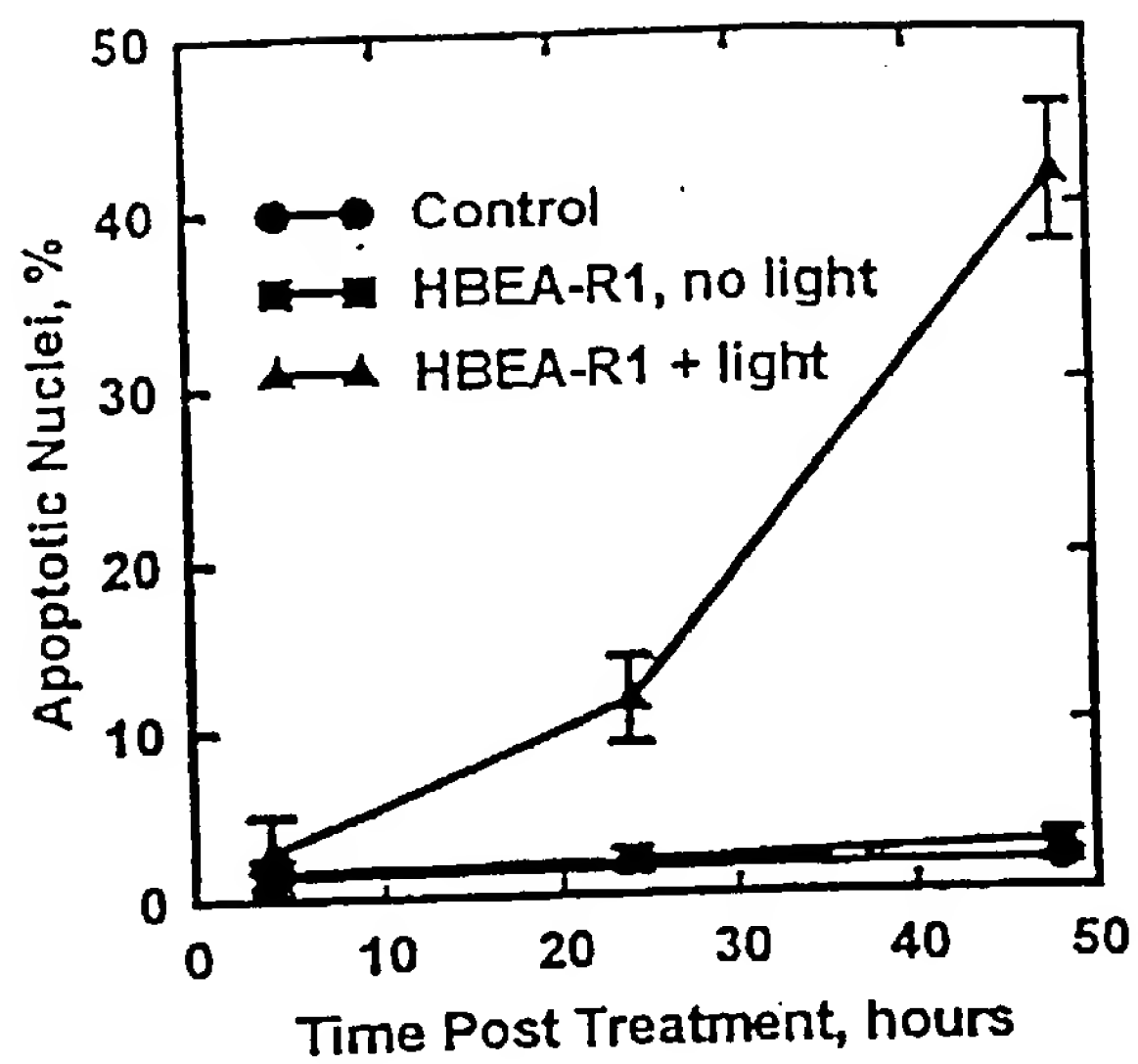


Figure 11:

HBEA-R1 - Induced Apoptosis in EMT6/Ed Cells



HBBA-R2 - Induced Apoptosis in EMT6/Ed Cells

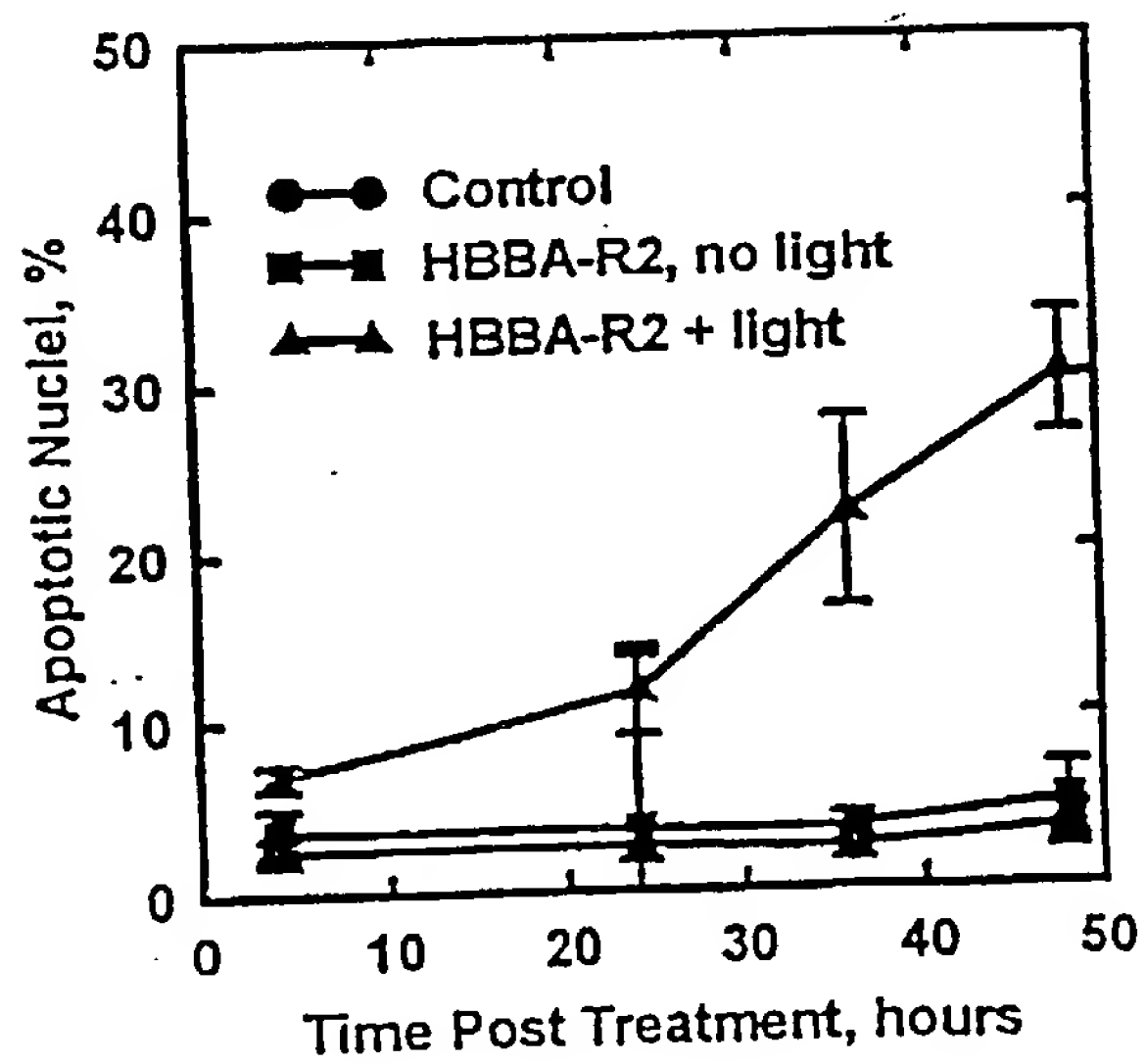
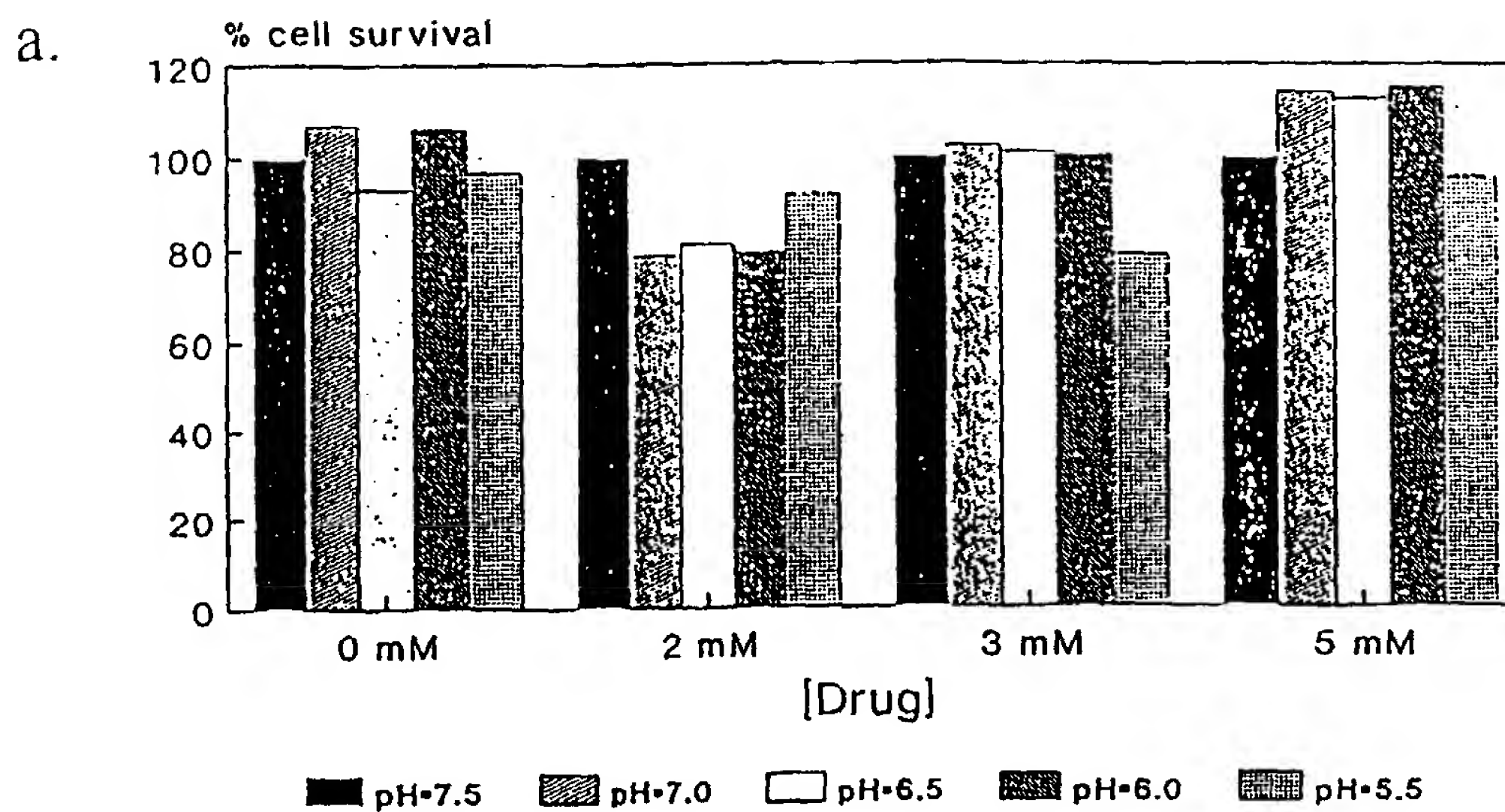


Figure 12.

pH Effect on HA Subjected to 630 nm light



b.

pH Effects on HA Dark Toxicity

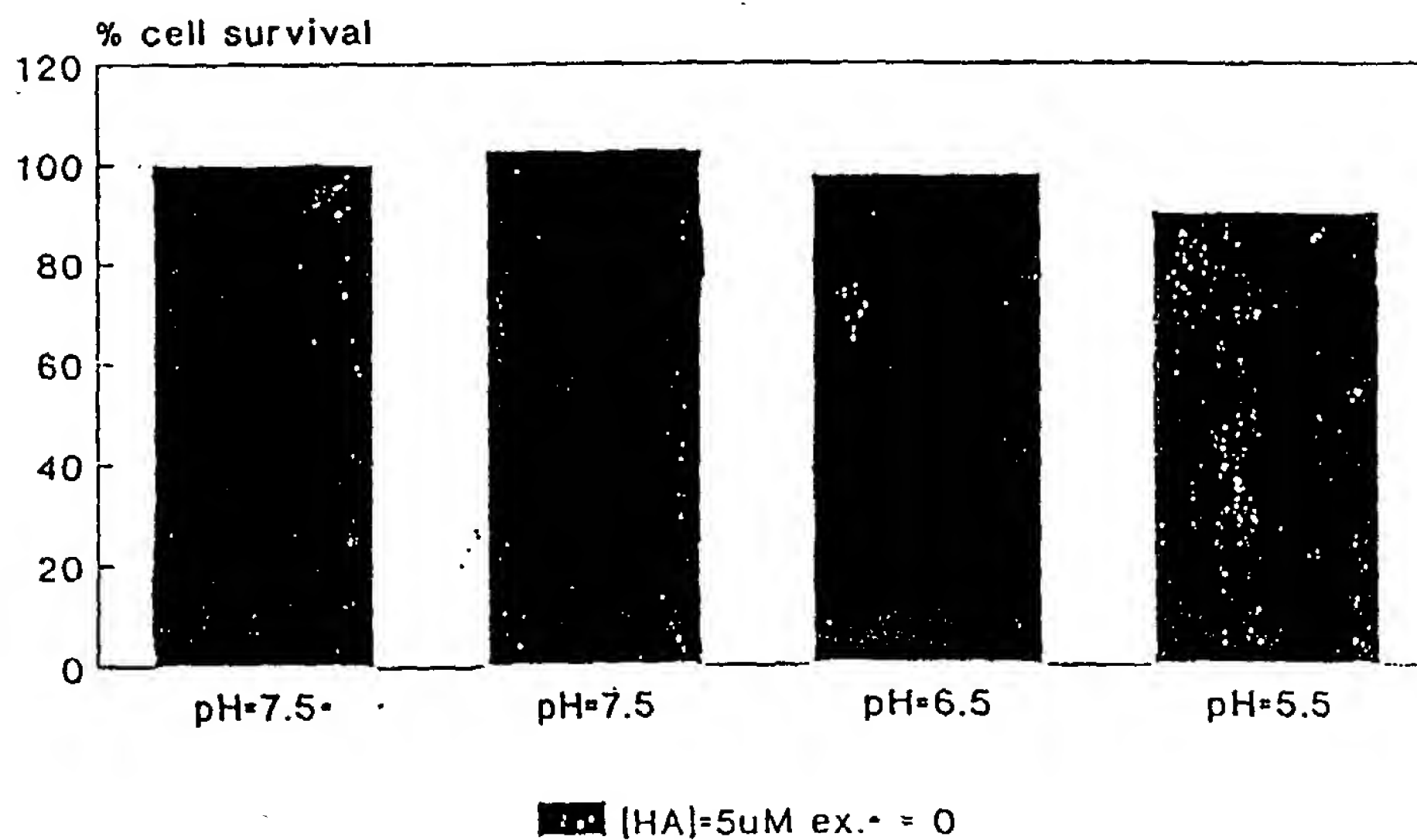
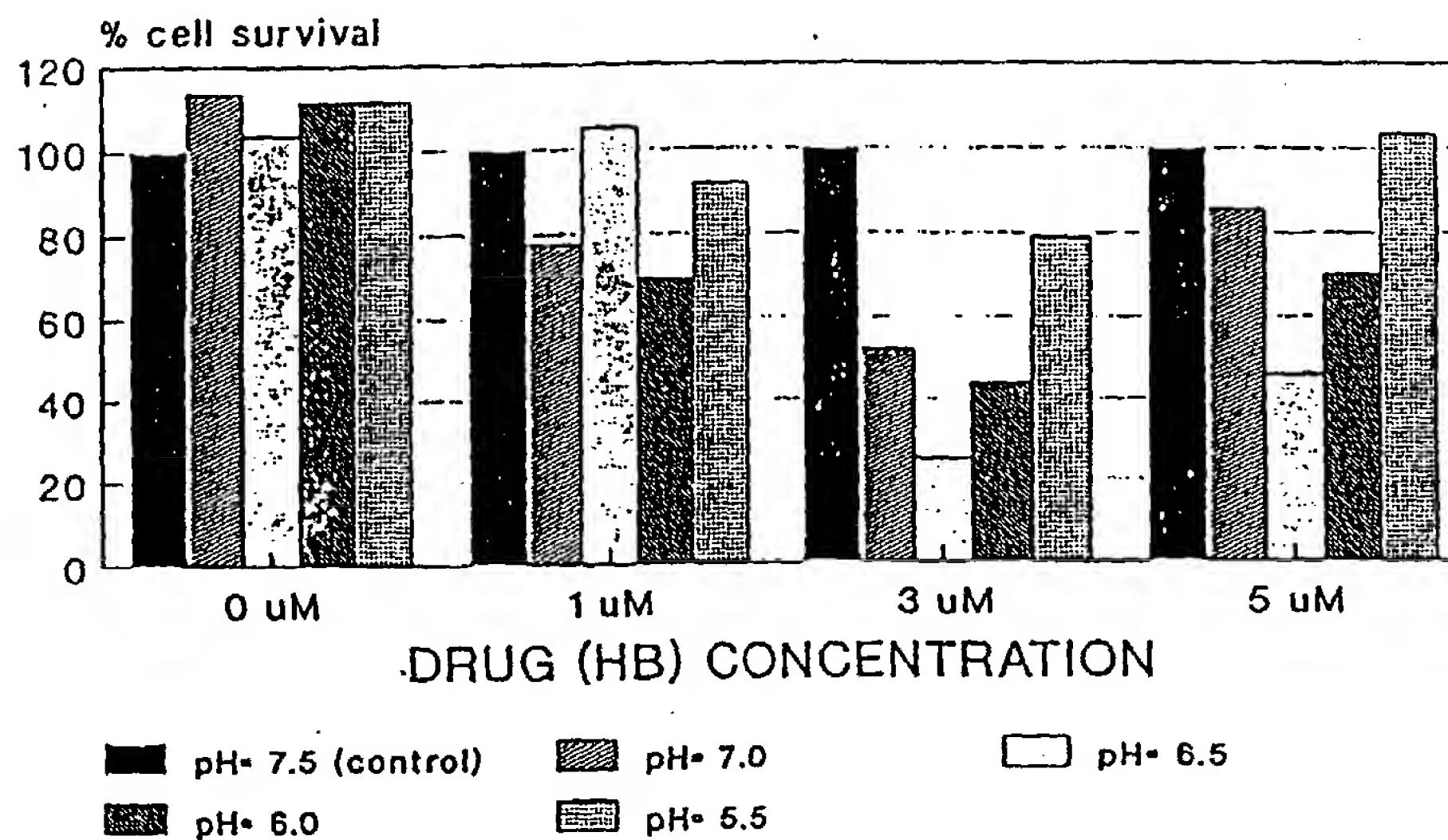


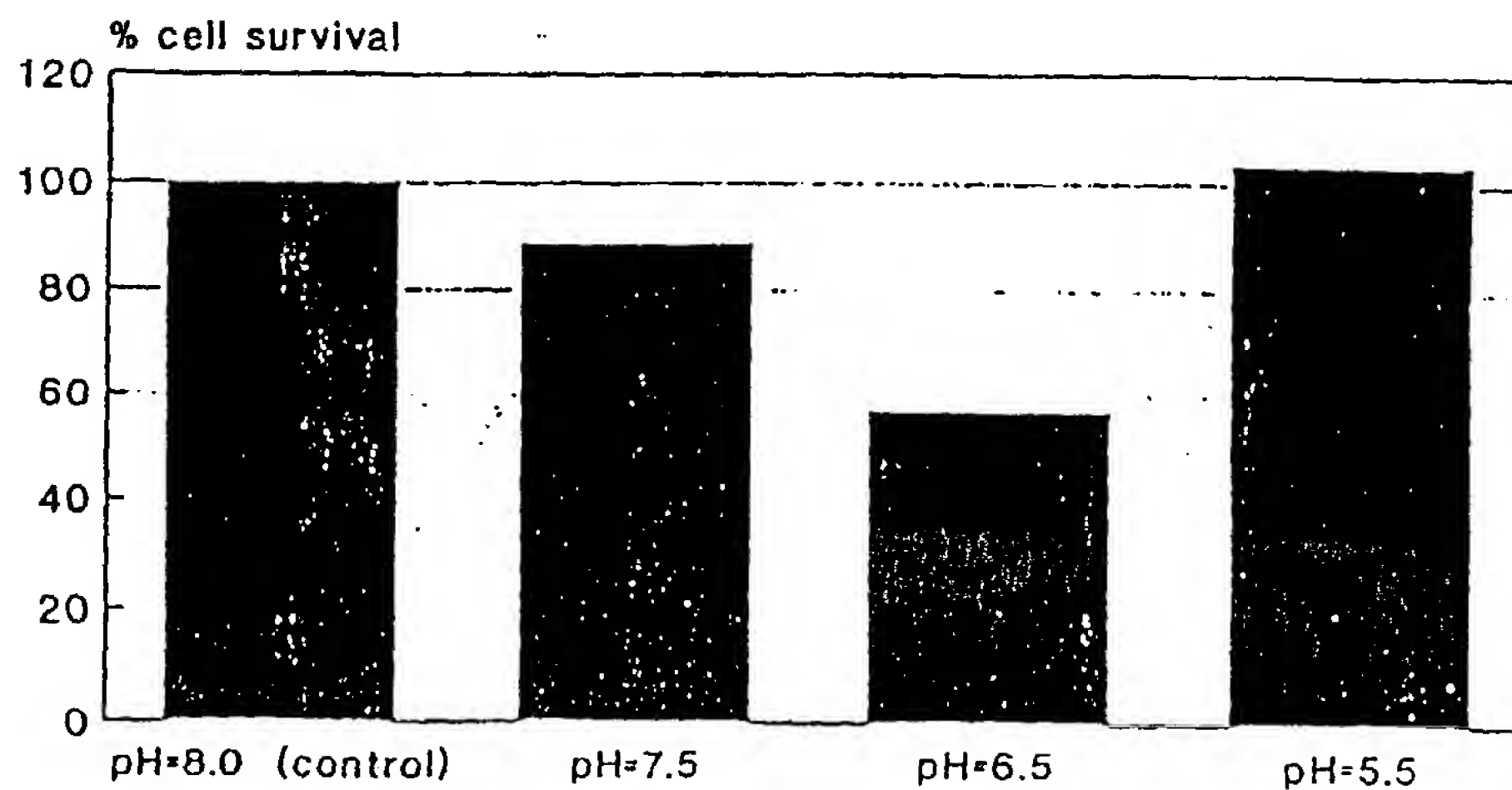
Figure 12. (CONT)

c. pH Effects on HB
Light @ 630 nm



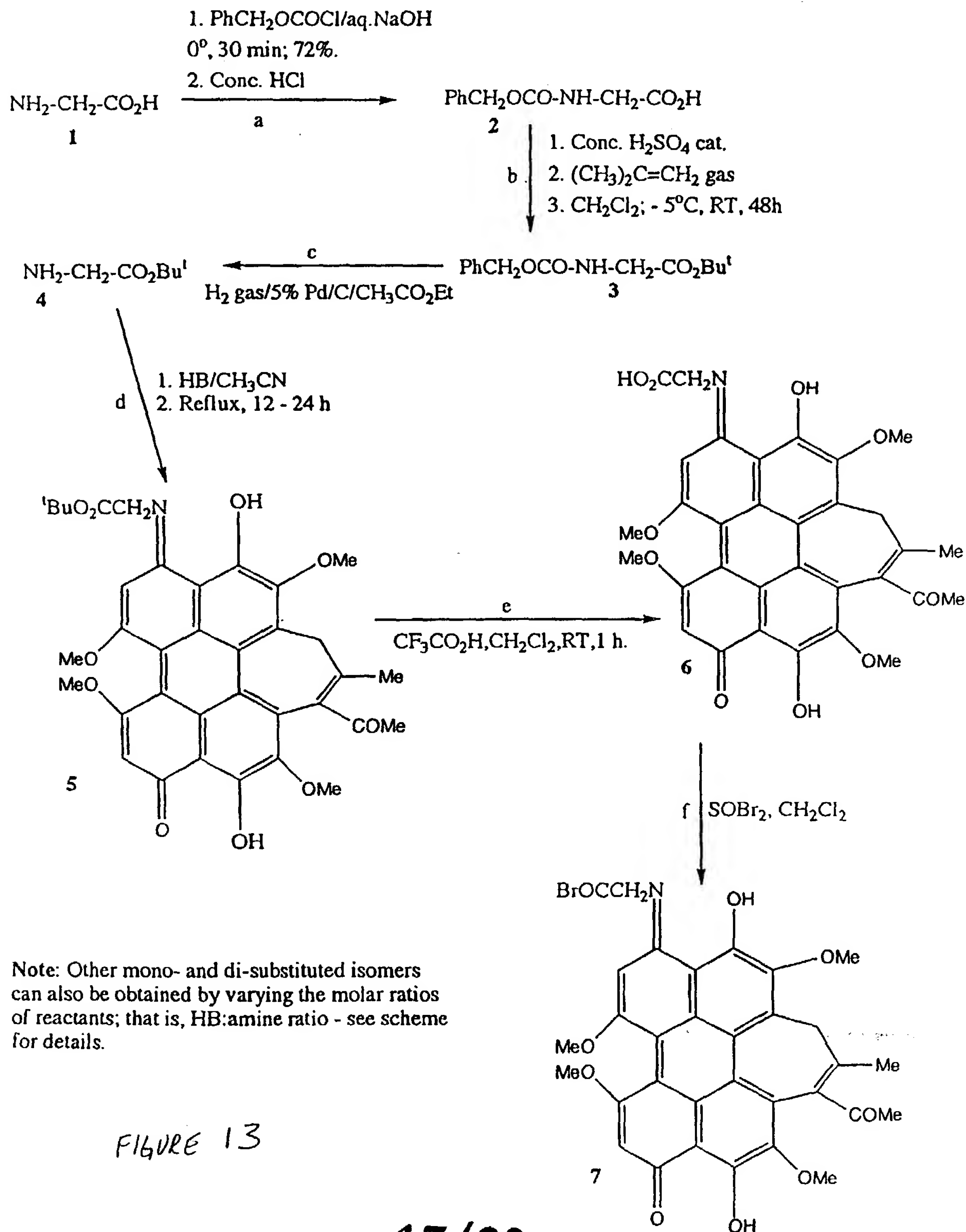
d.

pH Effects on HB
Dark Toxicity



[HB]-5uM

SYNTHESIS OF AMINO ACID DERIVATIVES OF HYPOCRELLIN B



FURTHER AMINO DERIVATIVES OF HYPOCRELLIN B

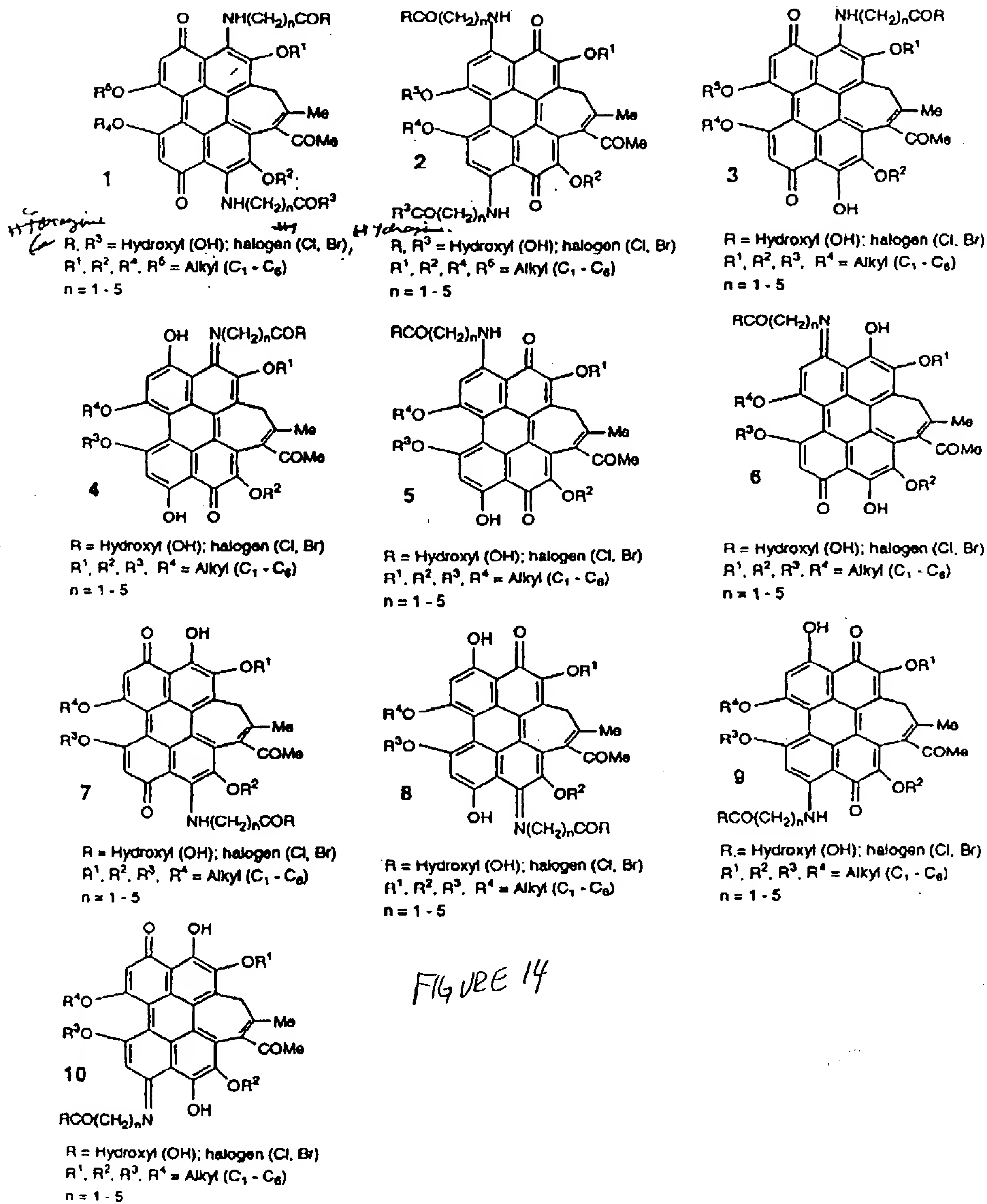
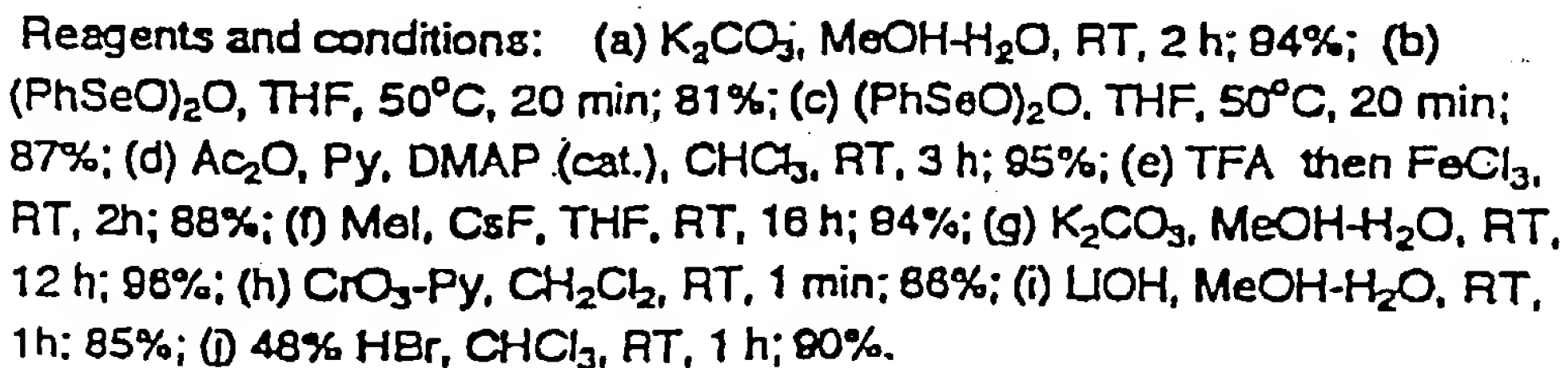


FIGURE 14

Scheme 11



Scheme 1

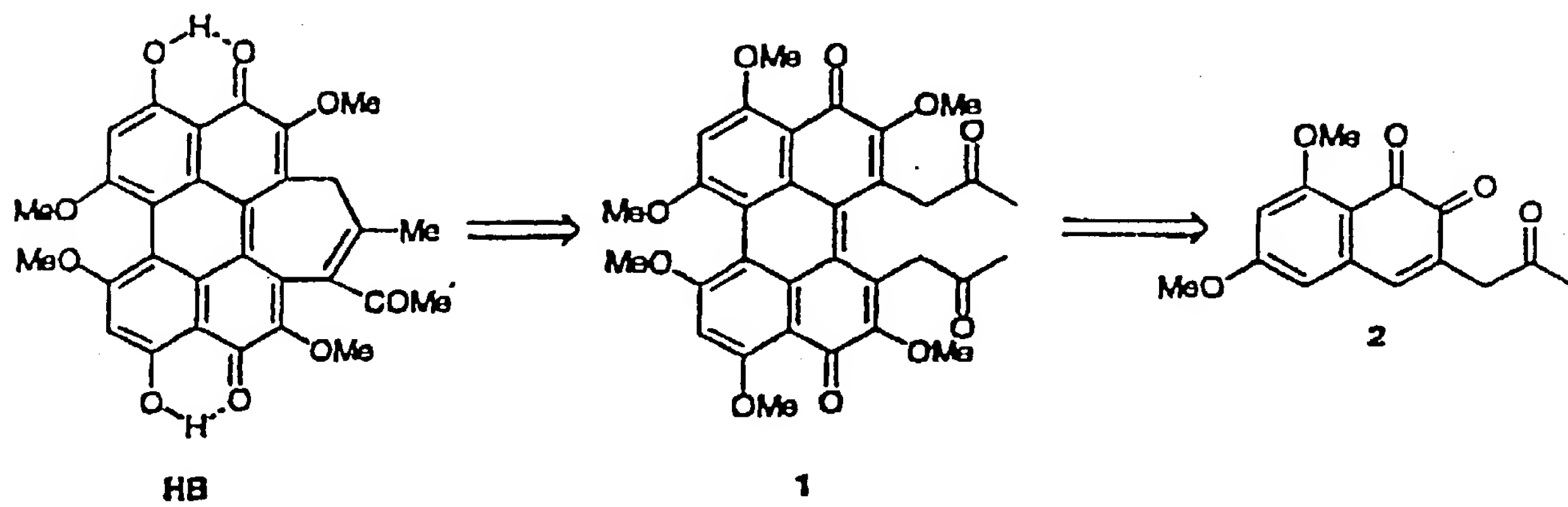


Figure 16